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(54) Title: METHOD FOR GENERATING RESISTANCE AGAINST CGMMV IN PLANTS, GENETIC CONSTRUCTS FOR USE IN SAID METHOD, AND CGMMV-RESISTANT PLANTS OBTAINED VIA SAID METHOD

(57) Abstract: The present invention relates to a method for generating resistance against Cucumber Green Mottle Mosaic Virus (CGMMV) in plants, in particular in plants susceptible to infection by CGMMV, such as *Cucurbitaceae* species, including melon, cucumber, watermelon and bottlegourd. In the invention, the plants are provided with replicase-mediated resistance against CGMMV. For this purpose, the plants are transformed with a polynucleotide sequence that: i) has been derived from the 129 kD sequence, the 57 kD sequence, or the 186 kD readthrough sequence of native CGMMV; ii) upon (at least) transformation into the plant and transcription into RNA - and usually also translation into the corresponding encoded protein - can provide the plant with resistance against CGMMV; but iii) does not encode any replicase activity. In particular, this will be a nucleotide sequence that encodes a defective variant of the replicase of CGMMV. The invention further relates to genetic constructs containing such a polynucleotide sequence suitable for use in the above method, and to CGMMV-resistant transgenic plants obtained via said method.



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Method for generating resistance against CGMMV in plants, genetic constructs for use in said method, and CGMMV-resistant plants obtained via said method.

The present invention relates to a method for generating resistance against Cucumber
5 Green Mottle Mosaic Virus (CGMMV) in plants, in particular in plants that are susceptible to infection by CGMMV, such as species of the *Cucurbitaceae* family.

The invention further relates to genetic constructs suitable for use in said method, and to CGMMV-resistant transgenic plants obtained via said method.

Methods of introducing DNA sequences into the genome of plants have been known
10 for many years and have been widely used to alter the properties of plants varieties. Such methods are among others *Agrobacterium*-mediated transformation (Horsch *et al.*, 1985; Rogers *et al.*, 1986), protoplast transformation using electroporation or other techniques to introduce naked DNA molecules into the plant cell (Shillito *et al.* 1985), and particle bombardment to introduce naked DNA molecules into plant cells or tissues (Christou *et al.*,
15 1994).

Among the most important applications of plant genetic engineering are those aimed at introducing resistance genes to a wide variety of plant pests and plant pathogens, such as bacteria, fungi, nematodes, insects and viruses. Many examples of virus resistance in a wide variety of plant species have been described over the last decades (Wilson *et al.*, 1993). The
20 various methods to obtain virus resistance in plants through the introduction of gene sequences are either based on the use of genes of plant origin; on the use of sequences/genes derived from the viral pathogen itself (so-called pathogen-derived resistance (Wilson *et al.*, 1993), or on the use of genes of yet different origin. Sequences originating from the viral genome can be either cloned or PCR-amplified DNA sequences obtained from the genome of
25 DNA viruses, such as geminiviruses (Kunik *et al.*, 1994) or the cDNA sequences obtained from the genomes of RNA viruses through the use of cDNA cloning or RT-PCR amplification.

Examples of sequences/genes of RNA viruses that have been successfully used in the engineering of virus resistance in plants include:

- 30 1. coat protein genes of tobamoviruses, cucumoviruses, potyviruses, potexviruses (Beachy *et al.*, 1990);
2. RNA-dependent RNA polymerase genes (replicase genes) of tobamoviruses, cucumoviruses, potyviruses (Anderson *et al.*, 1992; Donson *et al.*, 1993; Audy *et al.*, 1994);

3. nucleoprotein genes of tospoviruses (Goldbach and De Haan, 1993; Prins *et al.*, 1994; Vaira *et al.*, 1995);
4. movement protein genes of tobamoviruses and cucumoviruses (Cooper *et al.*, 1995).

5 Cucurbit Green Mottle Mosaic Virus (CGMMV) is a member of the tobamovirus group and infects plant species of the *Cucurbitaceae* family: melon (*Cucumis melo*), cucumber (*C. sativus*), watermelon (*Citrullus vulgaris*) and bottlegourd (*Lagenaria siceraria*), but not apparently *Cucurbita pepo* (squash, pumpkin, courgette). The host range of the virus is basically restricted to members of the *Cucurbitaceae*, with the exception that some but not
10 all strains also infect tobacco and/or the diagnostic species *Datura stramonium* and *Chenopodium amaranticolor* (Hollings *et al.*, 1975).

Several different strains can be distinguished serologically and by their response in *C. amaranticolor* and *D. stramonium* (Hollings *et al.*, 1975) The “type strain” was originally identified in Europe and does not normally cause fruit symptoms in cucumber. Another
15 European strain, called the *cucumber aucuba mosaic* strain, *cucumber virus 4* or *Cucumis virus 2A* causes fruit symptoms in cucumber. A number of strains are known from Japan. In watermelon, the *watermelon strain* causes serious disease, whereas the *Japanese cucumber strain* (also called Kyuri Green Mottle Mosaic Virus) and the *Yodo strain* cause fruit distortions in cucumber. The *CGMMV-C* strain from India is a pathogen on bottlegourd and
20 serious infections can cause complete crop losses.

In cucumber, CGMMV causes vein clearing, a light and dark green leaf mottle, leaf blistering and malformation and stunted growth, seriously affecting fruit yield. The East European isolates of the *aucuba mosaic strain* produces bright yellow leaf mottling and fruit discolouration.

25 CGMMV is transmitted through seed, but mostly through mechanical infection via the roots in contaminated soil, and through foliage contact and handling of plants (Hollings *et al.*, 1975). The virus particles are extremely stable and survive several months at normal temperatures. This stability combined with the very high infectivity through mechanical contact of the foliage is responsible for the economic importance of this virus, as even one or
30 a few infected plants in a cucumber greenhouse can eventually cause the infection and loss of the total crop. Also, infection may not only spread rapidly over a current crop, but also – due to the strong persistence of the virus – affect subsequent crops. Therefore, a CGMMV infection may require sterilisation of an entire greenhouse, as well as the use of sterile tools and materials.

The complete sequence of only one isolate of CGMMV has been determined (Ugaki *et al.*, 1991; Genbank accession numbers D12505 and D01188). This isolate “SH” had been found in infected watermelon plants in East Asia. Furthermore, the sequence of the coat protein gene of one other isolate (“W”) obtained from infected watermelon is known (Meshi *et al.*, 1983; Genbank accession numbers V01551 and J02054), as well as the sequence of the 29 kD movement protein gene of a watermelon strain (Saito *et al.*, 1988; Genbank accession number J04332). The nucleotide sequence of the CGMMV-SH isolate shows 55 to 56% identity with tobacco mosaic virus (TMV) and tobacco mild green mosaic virus (TMGMV), both other members of the tobamovirus group (Ugaki *et al.*, 1991).

As described by Ugaki *et al.*, the genome of CGMMV consists of a single-stranded RNA molecule coding for at least four open reading frames, encoding putative proteins of 186 kD, 129 kD, 29 kD and 17.3 kD, of which the 17.3 kD ORF is known to encode the coat protein. In this respect, Ugaki *et al.* state: “No CGMMV-encoded proteins except for the coat protein have yet been identified *in vivo*”.

The CGMMV genome is schematically shown in Figure 1. As can be seen therein, the ORF encoding the 186 kD protein starts at the same site as the ORF encoding the 129 kD protein, and adds a putative 57 kD polypeptide to the 129 kD ORF. The presence of this 57 kD protein alone has not been detected in infected plants. Instead, the 186 kD protein has been found, being the product of a read-through translation of the 129 kD and the 57 kD ORFs.

This 186 kD protein is thought to play a role in virus replication. Also, the 129 kD ORF is thought to encode a replicase function, whereas the 29 kD ORF is thought to encode a movement protein.

Hereinbelow, the nucleotide sequence corresponding to the ORF encoding the 129 kD protein will be referred to as “129 kD sequence”, the sequence corresponding to the 186 kD readthrough protein will be referred to as “186 kD sequence”, and the nucleotide sequence corresponding to the ORF encoding the 57 kD readthrough part will be referred to as “57 kD sequence”. These nucleotide sequences and the corresponding protein sequences are given in the sequence listings, as further described below.

Object of the invention was to provide a method for protecting plants, in particular plants susceptible to infection with CGMMV such as species of the *Cucurbitaceae* family, against infection with CGMMV, and in particular against infection with strains of CGMMV prevalent in Europe, such as the strains encountered in the cultivation of cucumbers in greenhouses.

Further objects were to provide means for use in said method, in particular a genetic construct that can be used for transforming plants or plant material so as to provide transgenic plants resistant against infection with CGMMV. Further objects of the invention will become clear from the description given hereinbelow.

5 For these purposes, applicant has investigated the symptomatology and the nucleotide sequence of the coat protein genes of 10 European strains of CGMMV, and compared these with the SH strain described by Ugaki *et al.*. A list of these strains, with their geographical origin and symptoms on cucumber, is given in Table 1.

10 Table 1. List of collected CGMMV-isolates with their geographical origin and symptoms on cucumber.

CGMMV isolate	geographical origin	symptoms on cucumber
1	Eastern Europe	vein clearing, mosaic
2	Eastern Europe	vein clearing, mosaic
3	IPO-DLO, the Netherlands	almost without symptoms
4	the Netherlands	weak leaf chlorosis
5	the Netherlands	weak leaf chlorosis
6	Proefstation Naaldwijk, the Netherlands	chlorosis
7	Rijk Zwaan, the Netherlands	chlorosis
8	Israël	chlorosis
9	Almería, Spain	chlorotic leaf spots
10	Almería, Spain	weak leaf chlorosis
CGMMV-SH	Japan	strong chlorotic leaf mosaic

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It was found that the sequences for the 10 European isolates are highly homologous (i.e. homology on the nucleotide level of 97%), and show about 90 % homology (on the nucleotide level) with the SH-isolate. The nucleotide sequences encoding the coat proteins of each of the isolates 1-10, as well as strain SH, are given in the sequence listings, as further described below. The corresponding phylogenetic tree is shown in Figure 2. This shows that the European isolates can be considered to constitute a subgroup of the CGMMV species.

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In the sequence listings:

- SEQ ID no.1 gives the nucleotide sequence encoding the 129 kD replicase protein of CGMMV isolate 4, with the ORF of the coat protein starting with the ATG codon at bp 523-525;
- SEQ ID no.2 gives the amino acid sequence of the 129 kD replicase protein of CGMMV isolate 4, with the ORF of the coat protein starting with the ATG codon at bp 523-525;
- SEQ ID no.3 gives the nucleotide sequence encoding the 57 kD protein of CGMMV isolate 4, with the ORF of the coat protein starting with the ATG codon at bp 523-525;
- SEQ ID no.4 gives the amino acid sequence of the 57 kD replicase protein of CGMMV isolate 4, with the ORF of the coat protein starting with the ATG codon at bp 523-525;
- 10 - SEQ ID no.5 gives the nucleotide sequence encoding the 186 kD readthrough protein of CGMMV isolate 4, with the ORF of the coat protein starting with the ATG codon at bp 523-525;
- SEQ ID no.6 gives the amino acid sequence of the 186 kD readthrough protein of CGMMV isolate 4, with the ORF of the coat protein starting with the ATG codon at bp 523-525;
- 15 - SEQ ID no.7 gives the nucleotide sequence encoding the coat protein of CGMMV isolate 1, with the ORF of the coat protein starting with the ATG codon at bp 523-525;
- SEQ ID no.8 gives the nucleotide sequence encoding the coat protein of CGMMV isolate 2, with the ORF of the coat protein starting with the ATG codon at bp 523-525;
- 20 - SEQ ID no.9 gives the nucleotide sequence encoding the coat protein of CGMMV isolate 3, with the ORF of the coat protein starting with the ATG codon at bp 523-525;
- SEQ ID no.10 gives the nucleotide sequence encoding the coat protein of CGMMV isolate 4, with the ORF of the coat protein starting with the ATG codon at bp 523-525;
- SEQ ID no.11 gives the nucleotide sequence encoding the coat protein of CGMMV isolate 5, with the ORF of the coat protein starting with the ATG codon at bp 523-525;
- 25 - SEQ ID no.12 gives the nucleotide sequence encoding the coat protein of CGMMV isolate 6, with the ORF of the coat protein starting with the ATG codon at bp 523-525;
- SEQ ID no.13 gives the nucleotide sequence encoding the coat protein of CGMMV isolate 7, with the ORF of the coat protein starting with the ATG codon at bp 523-525;
- 30 - SEQ ID no.14 gives the nucleotide sequence encoding the coat protein of CGMMV isolate 8, with the ORF of the coat protein starting with the ATG codon at bp 523-525;
- SEQ ID no.15 gives the nucleotide sequence encoding the coat protein of CGMMV isolate 9, with the ORF of the coat protein starting with the ATG codon at bp 523-525;

- SEQ ID no.16 gives the nucleotide sequence encoding the coat protein of CGMMV isolate 10, with the ORF of the coat protein starting with the ATG codon at bp 523-525;
- SEQ ID no.17 gives the nucleotide sequence encoding the 129 kD replicase protein of CGMMV isolate SH;
- 5 - SEQ ID no.18 gives the amino acid sequence of the 129 kD replicase protein of CGMMV isolate SH;
- SEQ ID no.19 gives the nucleotide sequence encoding the 57 kD protein of CGMMV isolate SH;
- SEQ ID no.20 gives the amino acid sequence of the 57 kD replicase protein of CGMMV isolate SH;
- 10 - SEQ ID no.21 gives the nucleotide sequence encoding the 186 kD readthrough protein of CGMMV isolate SH;
- SEQ ID no.22 gives the amino acid sequence of the 186 kD readthrough protein of CGMMV isolate SH;
- 15 - SEQ ID no.23 gives the nucleotide sequence encoding the coat protein of CGMMV isolate SH;
- SEQ ID's nos. 24-40 give the nucleotide sequences of the primers used in the Examples;
- SEQ ID's nos. 41-44 give the nucleotide sequences used in assembling the leader sequences used in the constructs described in the Examples;

20 In the above sequence listings, the nucleotide sequences given are DNA sequences, as the genetic constructs of the invention described below will usually contain or consist of a DNA. As CGMMV is an RNA virus, it will be clear to the skilled person that these sequences will occur in the virus as the corresponding RNA sequence (i.e. with U replacing T). Also, it will be clear to the skilled person that the nucleotide sequences given above may be followed

25 – both in the virus as well as in a construct of the invention – with a suitable termination codon, i.e. TAA/UAA, TAG/UAG or TGA/UGA (not shown).

Furthermore, as will be clear to the skilled person, the nucleotide sequence encoding the coat protein will usually start with an ATG codon. For example, in SEQ ID NOs 1 – 16, the nucleotide sequence encoding the coat protein starts at the ATG codon at base positions

30 523-525. (In the nucleotide sequence of SEQ ID NOs 1 – 16, the nucleotide sequence encoding the coat protein is preceded by another nucleotide sequence, e.g. encoding a movement protein. Accordingly, when hereinbelow reference is made to any nucleotide sequence of SEQ ID NOs 1 – 16, this also explicitly includes the nucleotide sequence starting at the ATG codon at base positions 523-525 of these SEQ ID's).

A particular purpose of the invention is therefore to provide a method that can provide plants with resistance against all these strains simultaneously, and more in particular a type of resistance that is agronomically useful, i.e. that can be used to protect (crops of) plants that are cultivated under circumstances wherein the high infectivity and persistence of CGMMV can be a major problem, such as the cultivation of cucumbers in greenhouses.

In the invention, this problem is generally solved by transforming a plant with a polynucleotide sequence (e.g. as part of a genetic construct) that can provide the plant with so-called "replicase-mediated" resistance against CGMMV. In particular, this will be a polynucleotide sequence that

- i) has been derived from the 129 kD sequence, the 57 kD sequence, or the 186 kD readthrough sequence of native CGMMV;
- ii) upon (at least) transformation into the plant and transcription into RNA - and usually also translation into the corresponding encoded protein - can provide the plant with resistance against CGMMV; but
- iii) does not encode any replicase activity.

In particular, such a polynucleotide sequence can encode a polypeptide or protein that is capable of providing a plant with resistance against GCMMV, but that by itself has no replicase activity, for instance due to one or more alterations in its amino acid sequence, compared to the amino acid sequence encoded by the 129 kD sequence, 57 kD sequence, and/or 186 kD readthrough sequence of native CGMMV.

However, according to one specific embodiment of the invention, the polynucleotide sequence may also comprise, or even consist of, the native 57 kD sequence.

In a first aspect, the invention therefore relates to a method for generating resistance in a plant against CGMMV, said method comprising at least the step of transforming said plant with a polynucleotide encoding a defective variant of the replicase gene of CGMMV.

In a further aspect, the invention also relates to a method for providing a transgenic plant and/or plant cell that is resistant against infection with CGMMV, comprising at least the step of transforming said plant or plant cell with a polynucleotide sequence encoding a defective variant of the replicase gene of CGMMV.

In another aspect, the invention also relates to a genetic construct suitable for transforming a plant, said construct at least comprising a polynucleotide sequence encoding a defective variant of the replicase gene of CGMMV, and optionally further elements of genetic constructs known per se. The invention also relates to a plant, plant cell and/or plant material that has been transformed with a genetic construct of the invention.

The invention also relates to transgenic plants that contain a polynucleotide sequence encoding a defective variant of the replicase gene of CGMMV, and/or that have been provided with resistance against infection with CGMMV by the method of the invention.

In the context of the invention, by the "*replicase gene of CGMMV*" is meant the native 129 kD sequence, the native 57 kD sequence, and/or the combined native 186 kD "readthrough" product of the native 129 kD and native 57 kD sequences.

By a "*native*" sequence is meant any RNA sequence that naturally occurs in CGMMV, including all isolates and strains thereof, as well as any DNA sequence that corresponds to these naturally occurring RNA sequences. Examples of such native sequences are the 129 kD nucleotide sequences given in SEQ. ID no.1 and SEQ. ID no.17, the 57 kD nucleotide sequences given in SEQ. ID no.3 and SEQ. ID no.19 and the 186 kD nucleotide sequences given in SEQ. ID no.5 and SEQ. ID no.21. It will be clear to the skilled person that there may be (further) naturally occurring variants of the RNA sequence from which the DNA sequences in the sequence listings were derived, and these (and the DNA sequences corresponding thereto) are also included within the term "*native sequence*".

By "*a polynucleotide sequence encoding a defective variant of the replicase gene of CGMMV*" in its broadest sense is meant a polynucleotide sequence that

- i) upon (at least) transformation into a plant and transcription into RNA generates resistance against infection with CGMMV in said plant; and
- ii) upon (at least) transformation into a plant and transcription into RNA does not lead to generation of (any) replicase activity in said plant (or at least -when it does lead to expression of some replicase activity - leads to expression of a replicase activity that is severely reduced compared to expression of the native gene encoding CGMMV replicase).

Herein, the terms "*plant*", "*transformed plant*" and/or "*transgenic plant*" include all parts or tissues of such a plant, including but not limited to individual cells of such a plant. These terms also includes material of or for such a plant, such as material that can be regenerated into a (mature) plant, including but not limited to protoplasts and/or callus tissue, or material that can be cultivated into a mature plant, such as cultivation material.

The plant is preferably a plant that is susceptible to infection with CGMMV, more preferably a plant belonging to the *Cucurbitaceae* family, such as melon (*Cucumis melo*), cucumber (*C.sativus*), watermelon (*Citrullus vulgaris*) and bottlegourd (*Lagenaria siceraria*).

Included within the term "CGMMV" are all known strains thereof, including those prevalent in Europe and Asia. In particular, the method of the invention can be used to protect

plants against strains of GCMMV prevalent in Europe (including Israel), such as those which are a problem in the cultivation of melons and in particular cucumbers in greenhouses, although the invention is not limited thereto.

In doing so, a major advantage of the invention is that it can provide protection against several, and preferably all, (such) strains of CGMMV simultaneously. Another advantage of the invention is that it provides "absolute" protection against CGMMV, which means that - upon expression of a polynucleotide sequence encoding a defective replicase in a plant - essentially no viral particles can be detected in the transformed plant (material). The method of the invention therefore does not lead to a deferral or slowing down of the onset of symptoms, as may occur when so-called "coat protein-mediated" resistance is used. Also, the method of the invention leads to a high level of resistance, and may also have the advantage of a favorable temperature effect.

Usually, the "*nucleotide sequence encoding a defective variant of the replicase gene of CGMMV*" will be a nucleotide sequence in which - compared to a nucleotide sequence encoding the corresponding native replicase of CGMMV - one or more nucleotides have been added, replaced and/or removed. In particular, the "*nucleotide sequence encoding a defective variant of the replicase gene of CGMMV*" may be a nucleotide sequence that comprises, and preferably consists of:

- a nucleotide sequence corresponding to the native 129 kD sequence in which - compared to said native sequence - one or more nucleotides have been added, replaced and/or removed;
- a nucleotide sequence corresponding to the native 186 kD sequence in which - compared to said native sequence - one or more nucleotides have been added, replaced and/or removed, e.g. in the part of the native 186 kD sequence corresponding to the 129 kD sequence, to the 57 kD sequence, or both;
- a nucleotide sequence corresponding to the native 57 kD sequence;
- a nucleotide sequence corresponding to the native 57 kD sequence in which - compared to said native nucleotide - one or more nucleotides have been added, replaced and/or removed;

such that said nucleotide sequence is capable - upon (at least) transformation into a plant and transcription into RNA - to confer to said plant resistance against infection with CGMMV, and such that said nucleotide sequence - upon (at least) transformation into a plant and transcription into RNA - is not capable of generating of (any) replicase activity in said plant.

Usually, the "*nucleotide sequence encoding the defective variant of the replicase gene of CGMMV*" will encode a protein or polypeptide, more specifically a protein or polypeptide that:

- 1) upon being expressed in a plant is capable of generating resistance against CGMMV in
5 said plant; and
- 2) upon being expressed in a plant has no replicase activity (or -when it has some replicase activity- has severely reduced replicase activity compared to the native CGMMV replicase).

Such a protein or polypeptide will be generally referred to hereinbelow as "*defective replicase*"; and a polynucleotide sequence encoding such a protein or polypeptide will be
10 referred to as a "*polynucleotide sequence encoding a defective replicase*".

Usually, the defective replicase will be a derivative - such as an analog, homolog, variant, mutant, part, fragment or combination of two or more such parts or fragments, etc.- of the amino acid sequence encoded by the native 129 kD sequence, the native 186 kD sequence
15 and/or the native 57 kD sequence, in which -compared to the amino acid sequence encoded by the corresponding native sequence- one or more amino acids have been added, replaced or removed, preferably replaced or removed, more preferably removed, leading to loss of replicase activity (or at least an inability to generate replicase activity when expressed in the plant).

20 In particular, the defective replicase may be a protein or polypeptide that comprises, and preferably consists of:

- an amino acid sequence corresponding to the amino acid sequence encoded by the native 129 kD sequence, in which - compared to said native sequence - one or more amino acids have been added, replaced or removed, preferably replaced or removed,
25 more preferably removed;
- an amino acid sequence corresponding to the amino acid sequence encoded by the native 186 kD sequence, in which - compared to said native sequence - one or more amino acids have been added, replaced or removed, preferably replaced or removed, more preferably removed, leading to loss of replicase activity;
- 30 - an amino acid sequence corresponding to the amino acid sequence encoded by the native 57 kD sequence;
- an amino acid sequence corresponding to the amino acid sequence encoded by the native 57 kD sequence, in which - compared to said native sequence - one or more

amino acids have been added, replaced or removed, preferably replaced or removed, more preferably removed;

or any combination thereof, provided that the resulting protein or polypeptide shows no replicase activity, but is still capable - upon expression in a plant - to generate resistance against CGMMV in said plant.

More in particular, the defective replicase may be a protein or polypeptide that comprises, and preferably consists of:

- an amino acid sequence corresponding to a part or fragment of the amino acid sequence encoded by the native 129 kD sequence, or to a combination of two or more such parts or fragments;
- an amino acid sequence corresponding to a part or fragment of the amino acid sequence encoded by the native 186 kD sequence, or to a combination of two or more such parts or fragments; or
- an amino acid sequence corresponding to the amino acid sequence encoded by the native 57 kD sequence.

such that the resulting protein or polypeptide shows no replicase activity, but is still capable - upon expression in a plant - to generate resistance against CGMMV in said plant.

An amino acid sequence "*corresponding to a part or fragment of the amino acid sequence encoded by the native 186 kD sequence, or to a combination of two or more such parts or fragments*" may for instance comprise: i) at least one part or fragment of the amino acid sequence encoded by the native 129 kD sequence combined with at least one part or fragment of the amino acid sequence encoded by the native 57 kD sequence (which combination of parts or fragments may or may not correspond to a contiguous amino acid sequence encoded by the native 186 kD sequence); ii) at least one part or fragment of the amino acid sequence encoded by the native 129 kD sequence combined with the full amino acid sequence encoded by the native 57 kD sequence, and/or iii) at least one part or fragment of the amino acid sequence encoded by the full native 129 kD sequence combined with at least one part or fragment of the amino acid sequence encoded by the native 57 kD sequence.

It is known, however, that expression in a plant of a nucleotide sequence encoding the full 129 kD sequence of the native replicase usually does not provide resistance against infection with CGMMV, but may even - upon infection of the plant - promote or facilitate multiplication of the virus. Therefore, in one embodiment, the invention does not comprise the expression in a plant of said replicase, nor the use of a polynucleotide sequence encoding such a replicase.

Even more preferably, the defective replicase is a protein or polypeptide that consists of:

- an amino acid sequence corresponding to a part or fragment of the amino acid sequence encoded by the native 129 kD sequence, or a combination of two or more such parts or fragments; such that the resulting protein or polypeptide shows no replicase activity, but is still capable, upon expression in a plant, to generate resistance against CGMMV in said plant; or
- an amino acid sequence corresponding to the amino acid sequence encoded by the native 57 kD sequence.

Any such parts or fragments may also contain one or more further amino acid substitutions, insertions or deletions compared to the native sequence, but this is not preferred.

Most preferably, the defective replicase is a so-called “truncated replicase”, i.e. an amino acid sequence corresponding to the amino acid sequence encoded either by the 129 kD sequence and/or by the 186 kD sequence, from which - compared to the native amino acid sequence - one or more amino acid residues are lacking at the carboxyl-terminus, such that the resulting protein or polypeptide shows no replicase activity, but is still capable - upon expression in a plant - to generate resistance against CGMMV in said plant. (In case of a truncated replicase based upon the 186 kD sequence, this usually means that the resulting protein will contain the full amino acid sequence of the 129 kD sequence, as well as part of the amino acid sequence of the 57 kD sequence (i.e. that is contiguous to the 129 kD sequence in the amino acid sequence encoded by native 186 kD sequence), with one or more amino acids lacking at the carboxy-terminus of the 57 kD part, although the invention in its broadest sense is not limited thereto).

The polynucleotide sequence that encodes such a truncated replicase may either comprise, or preferably consist of, the full native 129 kD sequence or 186 kD sequence, respectively, in which a stopcodon has been introduced at a desired site, or a polynucleotide sequence from which – compared to the full native 129 kD sequence or 186 kD sequence, respectively – one or more codons coding for the carboxy-terminal amino acid residues have been removed, i.e. starting from the 3' end of the native sequence(s).

As mentioned below, preferably a stopcodon is introduced in to the native sequence, in particular in the so-called GDD motif or in the P-loop. Examples thereof are the polynucleotide sequences comprised in the vectors shown in Figures 3-8, and as described in the Experimental Part.

Again, any such truncated replicase may also contain one or more amino acid substitutions, insertions or deletions compared to the native sequence, but this is not preferred.

As mentioned above, (the polynucleotide sequence encoding) the defective replicase is such that - after expression in a plant or plant cell - it is still capable of generating resistance against CGMMV in said plant. Usually, this means that the defective replicase will have at least one biological function that allows the defective replicase to protect the plant against CGMMV infection, such as for example down-regulation of viral replication or interference with the replication of the wild-type CGMMV, for instance by competing with wild-type virus for the replication machinery in the plant (cell). It will be clear that in order to achieve such a biological function, the defective replicase must usually have a certain minimal level of amino acid similarity with the amino acid sequences encoded by the native 129 kD, 186 kD and/or 57 kD sequences. In so far as the defective replicase is similar to the corresponding native amino acid sequence, this may be because it contains – on the corresponding amino acid positions - the same amino acid residues as the native amino acid sequence, or amino acid residues comparable thereto. The latter will usually comprise so-called “conservative” amino acid substitutions, for instance involving replacing a given acidic or basic amino acid residue by another acidic or basic amino acid residu.

However, there will also be differences in amino acid sequence between the defective replicase and the native replicase (i.e. the 129 kD, 186 kD or 57 kD protein), such that the defective replicase will no longer provide replicase activity. The skilled person will be able to select appropriate alterations to the amino acid sequence of the native replicase. As will be clear to the skilled person, a single (amino acid or nucleotide) alteration may be sufficient, or two or more such alterations may be required, dependant upon the position and nature of the alteration(s) compared to the amino acid sequence of the native replicase.

Whether a given polynucleotide sequence encodes a defective replicase according to the invention – or at least is capable of protecting a plant against infection with CGMMV – can simply be tested by transforming a plant, plant cell or plant material with a construct containing said polynucleotide sequence, and then exposing the plant, plant cell, plant material, and/or a mature plant generated therefrom, to CGMMV under conditions such that infection may occur. It can then be easily determined whether the polynucleotide sequence/construct is capable of protecting the plant, i.e. by suitably determining the presence of the virus, or simply by the presence or absence of symptoms of CGMMV-infection.

In general, as a minimum, when the defective replicase contains any amino acid substitutions or insertions, it will have an amino acid homology (i.e. identity on corresponding

position) with the corresponding native replicase protein of at least 80%, preferably at least 90%, more preferably at least 95%, with amino acid deletions not being taken into account, and a single amino acid insertion being counted as a single alteration.

In general, as a minimum, when the defective replicase contains one or more amino acid deletions, it will usually contain at least 30%, preferably at least 50%, more preferably at least 70%, and usually 80-90%, and may even contain as much as 95-99%, of the amino acid sequence of the corresponding native replicase protein, with any amino acid insertions or substitutions not being taken into account.

A truncated replicase based upon the 129 kD sequence will usually contain at least 50%, preferably at least 70%, and may contain as much as 80-95%, of the amino acid sequence of the native replicase. A truncated replicase based upon the 186 kD sequence may contain the full 129 kD protein followed by one or more amino acids from the 57 kD sequence, and usually contains the full 129 kD sequence followed by 1-95%, preferably 5-50%, of the 57 kD sequence.

The differences in amino acid sequence mentioned above can be differences compared to any of the amino acid sequences given in SEQ ID's 2, 4, 6 and/or 18, 20, 22, and/or compared to any naturally occurring variant of these amino acid sequences. These differences are at least such that the resulting protein does not correspond to a naturally occurring/native protein (including those given in SEQ ID's 2, 4, 6 and/or 18, 20, 22).

The polynucleotide sequences used in the invention are such that they encode the above defective replicases. For this purpose, they may contain the same codons as in the corresponding positions on the native 129 kD, 186 kD and/or 57 kD sequence, or codons equivalent thereto due to the degeneracy of the genetic code.

The polynucleotide sequence encoding the defective replicase can be provided in a manner known per se, for instance starting from the known sequence of the native 129 kD, 57 kD and/or 186 kD sequences, and/or from a nucleic acid that encodes said sequences. Usually, this will involve introducing one or more deletions, substitutions and/or insertions of one or more nucleotides, or even of one or more codons into, or compared to, the native sequence. Such deletions, substitutions and/or insertions will be collectively referred to hereinbelow as "*alterations*".

Accordingly, the polynucleotide sequence encoding the defective replicase may be a sequence that contains one or more such alterations compared to any of the nucleotide sequences given in SEQ ID's 1, 3, 5 and/or 17, 19, 21, and/or compared to any naturally occurring variant of these nucleotide sequences (including DNA sequences corresponding to

the RNA sequences as present in the virus). These differences are at least such that the protein encoded by the polynucleotide sequence does not correspond to a naturally occurring/native protein (including those given in SEQ ID's 2, 4, 6 and/or 16, 18, 22).

Furthermore, besides the alterations mentioned above, and compared to nucleotide sequences given in SEQ ID's 1, 3, 5 and/or 17, 19, 21 and/or compared to any naturally occurring variant of these nucleic acid sequences (including DNA sequences corresponding to the RNA sequences as present in the virus), the polynucleotide sequences may further contain one or more alterations that lead to a codon that encodes the same amino acid as the codon given for the corresponding position in SEQ ID's 1, 3, 5 and/or 17, 19, 21, and this may even lead to a fully or totally artificial and/or synthetic sequence. Also, compared to nucleotide sequences given in SEQ ID's 1, 3, 5 and/or 17, 19, 21 and/or compared to any naturally occurring variant of these nucleic acid sequences (including DNA sequences corresponding to the RNA sequences as present in the virus), the polynucleotide sequences may further contain one or more alterations that lead to a conservative amino acid substitution, i.e. as mentioned above.

Providing a polynucleotide sequence that contains the desired alterations will be within the skill of the artisan and can involve techniques such as nucleic acid synthesis using an automated nucleic acid synthesis technique; introduction of (point)mutations into a nucleic acid that comprises the native 129 kD, 57 kD, and/or 186 kD sequences; and/or using or suitably combining parts or fragments of the 129 kD, 57 kD and/or 186 kD sequences, or any combination thereof. Also, in providing such a polynucleotide sequence, the skilled person may take into account the degeneracy of the genetic code and/or conservative amino acid substitutions, as mentioned above.

In order to provide a polynucleotide sequence that encodes a truncated replicase as defined above, a technique involving the introduction of a stopcodon into the native sequence is particularly preferred.

A particularly preferred technique of introducing the above alterations – including stopcodons - involves the use of a PCR reaction, in which the desired alterations are introduced into the amplified sequence(s) by the use of modified primers, i.e. primers that contain a suitable “mismatch” compared to the template sequence, leading to the desired alteration in the amplified sequence. This PCR-based technique may also be used to introduce one or more restriction sites into the amplified sequence in order to facilitate the cloning of the amplification products into the desired transformation vectors.

As further described in the Experimental Part, this may involve a single PCR-reaction, but may also involve two or more PCR reactions, each leading to a part of intended final sequence encoding the defective replicase in which the primers (e.g. with the desired alteration) form the ends of the fragments. These fragments may then be combined, for instance to provide a polynucleotide sequence that comprises a combination of such fragments, and/or to reconstitute the full 129 kD, 57 kD and/or 186 kD sequence, now containing the desired alteration compared to the native sequence, such as a stopcodon.

The PCR-reactions and the further steps following amplification, such as combining/joining the amplified sequences, can be carried out in a manner known per se, for instance as described in the Experimental Part and/or using the techniques described in US-A-4,683,202; Saiki et al., Science 239 (1988), 487-491 or PCR Protocols, 1990, Academic Press, San Diego, California, USA.

As the template for the PCR-reaction, a nucleic sequence encoding the native 129 kD, 57 kD and/or 186 kD sequence can be used, such as a cDNA derived from the native RNA sequence, or a plasmid containing such a sequence, including those described in the Experimental Part. The template used may itself already contain one or more alterations, compared to the corresponding native sequence.

As mentioned above, a preferred alteration involves the introduction of a stopcodon into the native 129 kD or 186 kD sequence, such that – upon transformation into a plant – the polynucleotide sequence thus obtained causes expression of a truncated replicase. In particular, such a stopcodon may be introduced into a sequence corresponding to the native 129 kD sequence, more in particular to that part of the native sequence that corresponds to the so-called GDD-motiv or to the so-called P-loop.

The polynucleotide sequence encoding the defective replicase is preferably in the form of - e.g. forms part of and/or is incorporated within - a genetic construct. The genetic construct is preferably a construct suitable for the transformation of a plant, plant cell and/or plant material, such as a plasmid, cosmid or vector, including co-integration vectors or binary vectors. The genetic construct may be DNA or RNA, and is preferably dsDNA.

Such a construct may further contain all known elements for genetic constructs, and in particular for genetic constructs intended for the transformation of plants, as long as the presence thereof does not interfere with the CGMMV resistance to be provided by the polynucleotide sequence encoding the defective replicase. Some non-limiting examples of such elements include leader sequences, terminators, enhancers, integration factors, selection markers, reporter genes, etc., and suitable elements will be clear to the skilled person.

These further elements may or may not be derived from plants, and may or may not be homologous to the plant that is to be transformed with the construct of the invention (hereinbelow referred to as the “*target plant*”). For instance, the further elements may also have been derived from micro-organisms, viruses, etc., and may also be elements that are
5 natively associated with the CGMMV sequence, such as the native CGMMV leader sequence (5'-UTR sequence).

The nucleotide sequences encoding these further elements may have been isolated and/or derived from a naturally occurring source – for instance as cDNA – and/or from known available sources (such as available plasmids, etc.), and/or may have been provided
10 synthetically using known DNA synthesis techniques.

For instance, a construct of the invention will usually contain a suitable promoter operatively linked to the polynucleotide sequence encoding the defective replicase, e.g. such that it is capable of directing the expression of the polynucleotide sequence. Suitable promoters can be chosen from all known constitutive, inducible, tissue specific or other
15 promoters that can direct expression of a desired nucleotide sequence in a plant and/or in part of a plant, including specific tissues and/or individual cells of the plant. In particular, promoters are used that are suitable for use in species of the *Cucurbitaceae* family, such as cucumber.

A specifically preferred promoter is the plastocyanine-promoter. Use of the 35S
20 promoter is less preferred, as it may be less reliable in cucumber.

The terminator can be any terminator that is effective in plants. A particularly preferred terminator is the nos-3' terminator.

The selection marker can be any gene that can be used to select – under suitable conditions such as the use of a suitable selection medium- plants, plant material and plant
25 cells that contain –e.g. as the result of a succesful transformation - the genetic construct containing the marker. A particularly preferred selection marker is the *nptII*-gene, which can be selected for using kanamycin.

The construct of the invention further preferably contains a leader sequence. Any suitable leader sequence, including those of viral origin, can be used. Preferably, a leader
30 sequence essentially identical to the 5'untranslated (5'-UTR) region of the CGMMV genome is used. This may be derived from the viral RNA, or may be provided synthetically, e.g. as described in the Experimental Part.

Although not preferred, the invention also encompasses constructs that encode a fusion of a defective replicase as mentioned above, and at least one further amino acid

sequence, such as a protein or polypeptide, or a part or fragment thereof. Preferably, expression of a defective replicase as (part of) such a fusion does not detract from the desired biological activity (i.e. protection against infection with CGMMV).

The construct of the invention can be provided in a manner known per se, which generally involves techniques such as restricting and linking nucleic acids/nucleic acid sequences, for which reference is made to the standard handbooks, such as Sambrook et al, "Molecular Cloning: A Laboratory Manual" (2nd.ed.), Vols. 1-3, Cold Spring Harbor Laboratory (1989) of F. Ausubel et al, eds., "Current protocols in molecular biology", Green Publishing and Wiley Interscience, New York (1987).

According to one embodiment, the genetic construct is preferably (also) in a form that can be stably maintained or inherited in a micro-organism, in particular a bacterium, more in particular a bacterium that can be used to transform a plant or plant material, such as *Agrobacterium*. In a further aspect, the invention also relates to such a micro-organism, in particular a bacterium, more in particular a bacterium that can be used to transform a plant, such as *Agrobacterium*, that contains a genetic construct according to the invention.

The genetic construct can be transformed into the target plant, plant cell or plant material by any suitable transformation technique known per se, including transformation with *Agrobacterium*, transformation with "denuded" DNA, for instance through particle bombardment or transformation of protoplasts through electroporation or treatment with PEG.

Examples of suitable vectors systems for use with *Agrobacterium* are for instance binary vectors such as pBI121 and derivatives thereof; co-integration vectors such as pGV1500 and derivatives of pBR322. Suitable systems for transformation with denuded DNA include *E.coli*-vectors with high copy number, such as pUC-vectors and pBluescript II (SK+) vectors.

Upon transformation, the construct may for instance be incorporated into the genomic DNA of the plant, or it may be maintained/inherited independantly in the plant (cell).

In a further aspect, the invention therefore comprises a method in which a plant, plant cell or plant material is transformed with a genetic construct as described above.

This method may also comprise cultivating the transformed plant cell or plant material into a mature plant, and may also comprise sexually or asexually reproducing or multiplying the transformed plant (and/or the mature plant obtained from the transformed plant cell or plant material).

The invention therefore also relates to a plant, plant cell or plant material, that has been transformed with – or more generally contains- a genetic construct as described above.

Preferably, such a plant, plant cell or such plant material is resistant against infection with CGMMV as described herein.

The invention furthermore relates to cultivation material such as seed, tubers, roots, stalks, seedlings etc. for such a plant, as well as descendants of such a plant, obtained through sexual or asexual reproduction techniques. Such cultivation material and/or descendants most preferably still contain or have inherited the genetic construct of the invention, and more preferably also are resistant against infection with CGMMV as described herein.

The invention will now be illustrated by means of the following non-limiting Experimental Part and by means of the Figures, in which:

- Figure 1 is a schematic representation of the genome of CGMMV;
- Figure 2 gives a phylogenetic tree of CGMMV-coat protein (cp) for CGMMV-SH and the ten European isolates, using the method of J. Hein with weighted residue table.
- Figures 3 - 8 show examples of some preferred genetic constructs of the invention, i.e. those listed in Table 6 below.

Also, in the Experimental Part hereinbelow, enzymes, kits, etc. were usually used according to the instructions of the manufacturer and/or using well-established protocols, unless indicated otherwise.

Experimental Part

Example I: Cloning of the coat protein genes of 10 CGMMV-isolates

1. Collecting CGMMV isolates

To make use of coat protein-mediated protection (CPMP) strategy against CGMMV, it is necessary to clone the coat protein cistrons of the isolates, that are economically important.

As the only sequence information available for CGMMV is derived from watermelon strains from the Far East, it was first decided to collect CGMMV isolates of important cucumber culture areas in Europe and the Mediterranean area. Table 1 lists the isolates collected from various geographical areas. All isolates were propagated on cucumber, and infected leaf material was stored at -80°C . The symptoms obtained after infection of cucumber cv. *Hokus* are listed in Table 1.

2. Design of PCR primers

The possibility of sequence divergence among the various collected isolates, and between the isolates and the published sequences of CGMMV-SH and CGMMV-W exists. In

order to identify nucleotide regions with a high degree of sequence conservation, that could serve as a basis of PCR primer design, an alignment study was carried out on corresponding sequences of CGMMV-SH, CGMMV-W and of some other related members of the tobamovirus group: Sunn-Hemp Mosaic Virus (SHMV, a variant of TMV) and Pepper Mild Mottle Virus (PMMV). For this purpose, a region of 800 nucleotides just 5' of the coat protein cistron and a region of 170 nucleotides forming the far 3' of the viral genome were compared. In this sequence alignment, regions with sufficient sequence homology among all compared viruses were identified. Based on these sequences, sets of PCR primers were designed, which are listed in Table 2.

Table 2. Design of primers for the RT-PCR amplification of coat protein sequences of CGMMV-isolates.

primers	sequence	position on CGMMV-SH sequence
5' primers		
97G01	AGGTGTCAGTGGAGAACTCATTGA	5004
97G02	GGCGTTGTGGTTTGTGG	5210
97G03	CTGTAGGGGTGGTGCTACTGT	5248
3' primer		
97G18	GCCCATAGAACTTCAACGTC	6370

3. Amplification of the coat protein regions

From leaf material of cucumber plants infected with each of the 11 isolates described in Tabel 1, a total RNA extraction was prepared. Using each of the 5' primers listed in Table 2 in combination with 3' primer 97G18, reverse transcription of RNA and PCR amplification of cDNA with an annealing temperature of 55°C was established using a kit manufactured by Perkin Elmer Cetus. Especially in the reactions with the 5' primer 97G03 amplification products of the correct size were obtained for each of the 11 RNA samples. The PCR amplification products were directly cloned in T/A cloning vector pCR2.1 and introduced in *E.coli* strain INVαF'. For each of the RNA samples, the correct size of the cloned product (1.12 kb) was verified, and the clones were stored at -80°C. The amplification products of

CGMMV-isolates 1 to 10 cloned in pCR2.1 were designated pKG4301 to pKG4310, and the one of CGMMV-SH cloned in pCR2.1 was designated pKG4311.

4. Nucleotide sequence analysis of the coat protein cistrons

5 The sequences of the complete inserts of the plasmids pKG4301 to pKG4310 were determined by reading in both directions using m13 forward and m13 reverse sequencing primers. The sequence of the insert of pKG4311 was already known, as this plasmid contains a cDNA fragment of CGMMV-SH.

10 Sequence analysis confirmed, that in each case indeed the correct cDNA fragment of CGMMV had been obtained and cloned. With one exception, each amplified and cloned cDNA fragment consisted of 1123 base pairs, containing the CGMMV coat protein cistron and a large part of the CGMMV movement protein cistron.

15 The cloned sequences of all collected European isolates (isolates 1 to 10) are approximately 97% homologous among each other, but differ on average by 10% from the published sequence of CGMMV-SH. Comparison of each individual sequence revealed, that isolates 1 and 2, both from Eastern Europe, are extremely alike. The same very high degree of identity was found between both isolates from cucumber greenhouses in the Netherlands (isolates 4 and 5) and between both isolates obtained from the Almería area in Spain (isolates 9 and 10). None of the cDNA sequences was 100% identical to any of the other ones, but the differences in sequence are no more than a few nucleotides, and sometimes only one nucleotide in the coding region of the coat protein cistron. The Japanese isolate CGMMV-SH is clearly different from any of the European isolates.

5. Coat protein amino acid sequence analysis

25 Based on the nucleotide sequences of the Open Reading Frames (ORF) of the coat protein cistrons of the 10 isolates, the amino acid sequence could be deduced. In each of the analyzed sequences, the ORF consisted of a region of 486 nucleotides, coding for a protein of 161 amino acid residues. The predicted molecular mass of this protein is 17.3 kD, corresponding to earlier published results. The homology among the predicted protein sequences of the various isolates is as high as 98.1%. The only deviations are found for amino acid residue 19 (usually valine), residue 65 (mostly serine) and residue 84 (mostly leucine).

30 The sequence of the coat protein of the Japanese isolate CGMMV-SH only differs by 1 amino acid (residue 65) from the consensus sequence.

Example II: Cloning of the replicase gene of CGMMV

1. Strategy for replicase-mediated protection

5 By way of example, two approaches to replicase-mediated protection (RMP) against virus infections in plants were investigated.

One approach makes use of defective replicase genes in the form of truncated Open Reading Frames (ORF), in which the sequence downstream from the GDD motif had been truncated or altered through mutation.

10 The other approach makes use of the expression of the 'read-through' part of the replicase gene, i.e. the 57 kD sequence. It is thought that this ORF is not translated in the plant cell, but forms part of a larger 'read-through' ORF combining the coding regions of both the 129 kD replicase gene and the putative 57 kD protein gene, resulting in a protein of 189 kD. However, expressing merely the 57 kD protein ORF in plant cells may result in a
15 extremely strong resistance to infection by both virus particles and viral RNA, which also would be capable of resisting high temperatures, as well as high inoculum concentrations.

For either of these approaches, either the full-length CGMMV replicase gene must be cloned, or both constituting parts must be cloned separately.

2. Design of primers

20 Because of the high sequence homology of the coat protein genes of 11 CGMMV isolates it was assumed, that the sequences of the replicase genes of the various isolates would be also highly conserved. Based on the complete sequence of the CGMMV-SH isolate,
25 primers were designed for the PCR amplification of the 57 kD ORF and of the 129 kD ORF (Tables 3 and 4). The primers were designed such, that they contain restriction sites for the future cloning of the amplification products. The 5' primers contain an *NcoI*-site positioned such, that it will coincide with the ATG start codon of the amplified ORF. The 3' primers contain a *SacI*-site downstream from the stop codon.

Table 3. Design of primers for the LR-RT-PCR amplification of the 57 kD replicase sequence of CGMMV.

primers	sequence	position on CGMMV-SH sequence
5' primer		
98A88	CCATGGAGAATTCGCTGTATGTC C	3497
3' primer		
98A86	CGAGCTCTCGACTGACACCTTAC	5001

5 Table 4. Design of primers for the LR-RT-PCR amplification of the 129 kD replicase gene sequence of CGMMV.

primers	sequence	position on CGMMV-SH sequence
5' primers		
98A84	CCATGGCAAACATTAATGAAC	59
98A85	CAACCATGGCAAACATTAATG	56
3' primer		
98G63	TAACAGGGAGGAAAATATTACG	

10 3. Long-Range Reverse Transcriptase Polymerase Chain Reactions

From the known sequence of CGMMV-SH it was derived that the size of the 57 kD protein gene is 1.5 kb. Such a size is at the limit of the size range that can be amplified in a PCR with standard *Taq* polymerase. For the amplification of this cDNA fragment, and certainly for the amplification of the cDNA fragment for the 129 kD replicase gene, a
 15 different polymerase suitable for long range amplifications must be used. In these experiments, rTth DNA polymerase was used.

For direct amplification of cDNA fragments from total RNA extractions a RT-PCR kit is normally employed, combining in one reaction the activity of the Reverse Transcriptase (RT), producing a single cDNA strand complementary to the RNA template strand beginning

at one primer annealed to the 3' end of the RNA molecules, and the activity of the Polymerase, amplifying the thus produced single stranded cDNA molecule in a normal PCR fashion.

Because of the need to use long range polymerase, it was attempted to combine the RT with the long range polymerase to produce in one reaction large-size amplification products directly from total RNA extracts. This type of reaction was called a Long Range Reverse Transcriptase Polymerase Chain Reaction (LR-RT-PCR).

4. LR-RT-PCR amplification and cloning of the 57 kD protein gene

Using the primers listed in Table 3 and the LR-RT-PCR described above, a specific 1.5 kb amplification product was obtained from total RNA extracts of cucumber leaves infected with CGMMV-4. This isolate was chosen, as it originated from the Dutch cucumber greenhouse cultures, and would thus represent an economically important isolate. Because long range polymerases contain a 'proof reading' activity and do not leave A-additions on the amplification products, as does the *Taq* polymerase normally employed in PCR, direct cloning of the amplification products in a TA vector accomodating the A-additions was not possible. Therefore, the amplification products were briefly treated with *Taq* polymerase, resulting in the addition of A-overhangs on the amplified DNA molecules. These molecules could then easily be cloned in the TA vector pCR2.1, and transformed to *E.coli* MC1061. Clones with the correct insert size of 1.5 kb were stored at -80°C and are known as pKG4321.

5. Sequence analysis of the 57 kD protein gene

The nucleotide sequence of the cloned insert of pKG4321 was determined by double-stranded sequencing using m13 forward en m13 reverse primers and subsequent primer walking steps. The ORF coding for a putative 57 kD protein gene (SEQ ID no 3) showed 90% homology at the nucleotide level to the corresponding sequence of the Japanese isolate CGMMV-SH (SEQ ID no 19). The predicted amino acid sequence (SEQ ID no 4) shows a 98.2% homology to the one predicted by the CGMMV-SH sequence (SEQ ID no 20). The GDD motif characteristic for viral replicase genes resides at amino acid residues 364-366.

6. LR-RT-PCR amplification and cloning of the 129 kD replicase gene

Using the primers listed in Table 4 in a Long Range Reverse Transcriptase Polymerase Chain Reaction as described under 3 , one specific amplification product of 3.5 kb

representing the viral 129 kD replicase gene was obtained from total RNA of cucumber leaves infected with CGMMV isolate 4 (Table 1). Because long range polymerases contain a 'proof reading' activity and do not leave A-additions on the amplification products, as does the *Taq* polymerase normally employed in PCR, direct cloning of the amplification products in a TA vector accomodating the A-additions was not possible. Therefore, the amplification products were briefly treated with *Taq* polymerase, resulting in the addition of A-overhangs on the amplified DNA molecules. These molecules could easily be cloned in the TA vector pCR2.1, and transformed to *E.coli* MC1061. Clones with the correct insert size of 3.5 kb were stored at -80°C and are known as pKG4322.

7. Sequence analysis of the 129 kD protein gene

The nucleotide sequence of the amplification product cloned in pKG4322 was determined by double-stranded sequencing using m13 forward en m13 reverse primers, and a primer walking strategy. The ORF coding for the 129 kD replicase gene (SEQ ID no 1) showed 88% homology at the nucleotide level to the corresponding sequence of the Japanese isolate CGMMV-SH (SEQ ID no 17). The ORF of the Dutch cucumber greenhouse isolate codes for a replicase protein of 1144 amino acids, which is one amino acid in extra in comparison to the CGMMV-SH strain. The predicted amino acid sequence (SEQ ID no. 2) shows a 97.1% homology to the one predicted by the CGMMV-SH sequence (SEQ ID no 18).

Two GDD motifs are found at amino acid residues 256-258 and 540-542.

8. Site-directed mutagenesis of the 129 kD ORF

As explained above, one approach to obtain RMP in plant cells was to make use of replicase genes truncated either in the GDD motif, or truncated in the P-loop of the helicase domain. In order to create gene expression cassettes carrying such truncated genes, a site-directed mutagenesis approach was followed to introduce stop codons at the required positions in the ORF. To this end, several parts of the 129 kD replicase ORF were re-amplified from pKG4322 as a template using specifically designed primers that included unique restriction sites for future re-assembling of the thus amplified products, as well as the required mutations in the form of stop codons (Table 5). These stop codons should ensure the proper truncation of the translation of the protein. Several stop codons were designed one after the other in the three reading frames in these primers, thus ensuring an effective translation-deficient mutation.

Table 5. Design of primers for the site-directed mutagenesis of the 129 kD replicase gene of CGMMV.

primers	sequence	
98L99	GAGCTCGGATCCACTAGTAACGGC	
98L107	TAGAGCTCTTGAAGCTAAGCAAATTCCG	
98L108	TTCAAGAGCTCTAATCACCGAAGACAAAGGC	
98L102	GAATTATATCGATTATCTATCGGC	
98L103	GATAATCGATATAATTCTTCATCTGCC	
98L104	AACTAGTAATTGATGATCTGTTCAAGAAG	
98L105	AATTACTAGTTTCCGGAAGCAAGCAGCTCAG	
98L106	GCCCTCTAGATGCATGCTCGAG	

5 Using primers 98L103 and 98L104, a fragment from the downstream half of the 129 kD gene from the GDD motif up to the *Clal*-site was amplified, while simultaneously stop codons were introduced at the site of the GDD motif. This fragment cloned in TA vector pCR2.1 was called pKG4325.

10 Using primers 98L105 and 98L106, a fragment corresponding to the 5' half of the 129 kD gene up to the GDD motif was amplified, while simultaneously a stop codon was introduced at the site of the GDD motif. This fragment cloned in T/A vector pCR2.1 was called pKG4326.

15 Replacing an *XbaI-Clal* fragment of pKG4322 with the combined amplified products of pKG4325 and pKG4326 reconstitutes the full-length 129 kD replicase ORF of pKG4322 with stop codons introduced at the site of the GDD motif. This construct is named pKG 4329.

Using primers 98L99 and 98L107, a fragment at the far downstream end of the 129 kD gene from the P-loop to the end of the ORF was amplified, while simultaneously stop codons were introduced at the site of the P-loop. This fragment cloned in T/A vector pCR2.1 was called pKG4327.

20 Using primers 98L108 and 98L102, a fragment corresponding to a central part of the 129 kD gene from the GDD motif up to the P-loop was amplified, while simultaneously a stop codon was introduced at the site of the P-loop. This fragment cloned in T/A vector pCR2.1 was called pKG4328.

Replacing an *Bam*HI-*Cl*al fragment of pKG4322 with the combined amplified products of pKG4327 and pKG4328 reconstitutes the full-length 129 kD replicase ORF of pKG4322 with stop codons introduced at the site of the P-loop. This construct is named pKG4330.

Example III: Transformation of cucumber

1. Construction of a CGMMV-leader sequence

For optimal expression and stability of the replicase gene transcripts in plant cells, it was thought necessary to add a sequence identical to the 5' untranslated (5' UTR) region of the CGMMV genome upstream from the ORF sequence in the plant expression cassette. Because the 5' UTR of viral genomes contain highly repetitive RNA, this sequence could not be obtained by RT-PCR amplification, as no specific primers could be designed. Instead, a synthetic region identical to the 5' UTR of CGMMV-SH was assembled from the four oligonucleotide sequences:

97G40 (CTAGAGTTTTAATTTTTATAATTAAACAAA),
97G41 (TCAAAATTAAAAATATTAATTTGTTTGTTGTTG),
97G42 (CAACAACAACAACAACAATTTTAAAACAACAC) and
97G43 (TTGTTGTTTGTTAAAATTTTGTTGTGGTAC).

These oligonucleotides were designed such, that outside the sequence corresponding to the 5' UTR they contain restriction sites for *Xba*I and *Nco*I, thus facilitating further cloning. Adding the four oligonucleotides together will cause spontaneous assembling due to the design of extensive regions of overhang. Using these restriction sites, the assembled mixture was cloned in a plant expression vector containing an *Arabidopsis thaliana* plastocyanin promoter (Vorst *et al.*, 1993) and a *Agrobacterium tumefaciens* nopaline synthase terminator sequence (Depicker *et al.*, 1982) in a pUC19-derived plasmid. This expression cassette was called pKG1315. The complete expression cassette consisting of the plastocyanin promoter, the CGMMV leader sequence and the *nos* terminator was subsequently removed from pKG1315 using *Hind*III and *Eco*RI as restriction enzymes, and recloned in the corresponding restriction sites of:

- 1) an intermediate type *Agrobacterium* transformation vector for coïntegrate type vector systems containing an *nptII* selectable marker gene cassette to create pKG1575, and
- 2) an intermediate type *Agrobacterium* transformation vector for coïntegrate type vector systems containing no selectable marker gene to create pKG1110.

2. Construction of transformation vectors

The three cloned and modified replicase constructs of pKG4321, pKG4329 and pKG4330 were isolated from the plasmids by restriction with *Bam*HI (filled in with Klenow) and *Nco*I and ligated into the *Sac*I (filled in with Klenow) and *Nco*I sites of each of the two transformation vectors pKG1575 and pKG1110, resulting in a total of six transformation vectors, listed in Table 6.

Table 6. List of six transformation vectors for the expression in plants of parts of the CGMMV-replicase gene.

vector	vector type	modified CGMMV-replicase gene
pKG4331	intermediate type with nptII	57 kD ORF
pKG4332	intermediate type with nptII	129 kD ORF with stopcodon in GDD motif
pKG4333	intermediate type with nptII	129 kD ORF with stopcodon in P-loop
pKG4334	intermediate type	57 kD ORF
pKG4335	intermediate type	129 kD ORF with stopcodon in GDD motif
pKG4336	intermediate type	129 kD ORF with stopcodon in P-loop

3. Transformation of cucumber

The intermediate type transformation vectors pKG4331 and pKG4333 were introduced into *Agrobacterium tumefaciens* strain GV2260 by tri-parental mating.

Transconjugants which had incorporated the intermediate type vector into their Ti-plasmids through homologous recombination were selected on the basis of streptomycin and spectinomycin resistance and analyzed for the correct insertion of the vector.

Cucumber plants were transformed with these two strains of *Agrobacterium*, as well as with an *Agrobacterium* strain harbouring only the *nptII* selection marker, using published procedures. A number of transgenic cucumber plants were obtained. The plants were transferred to a greenhouse to flower and set seed. The seedlings germinating from these R1 seed were mechanically infected with CGMMV isolate 1-3 weeks post-inoculation, the plants were scored for symptoms of virus infection, as described in the assay for tolerance to virus infection set out in under 4. below.

4. Assay for tolerance to virus infection

The seedlings of transgenic cucumber germinating from these R1 seed were mechanically infected with CGMMV isolate 1. Fresh inoculum was prepared from a crude leaf extract of susceptible non-transgenic cucumber plants cv. Hokus pre-infected with this same isolate 3 weeks previously. Seedlings of non-transformed cucumber plants were used as controls in the assay. During 21 days post-inoculation the appearance of viral symptoms was scored visually every 2 days. In this assay, individual plants are scored as being tolerant when they remain free of visible symptoms for at least 7 days, and preferably more than 14 days, and more preferably more than 21 days post-inoculation.

Sixty-four independent transgenic lines were assayed, with 14 to 20 seedlings for each line. Control seedlings all became diseased within 9 days post-inoculation. A number of seedlings in seventeen of the transgenic lines showed clear absence of symptoms for a prolonged period of time, and remained free of symptoms after 21 days post-inoculation. Of some transgenic lines, the number of symptom-free plants corresponded to Mendelian segregation of a transgene present in a single locus. In one particular transgenic cucumber line 4 out of 14 seedlings remained symptom-free during the assay period, which may indicate that the tolerant phenotype corresponds to the homozygous state of a transgene present in one single locus, although, as mentioned above, the invention is not limited to a specific mechanism.

Hereinabove, the invention has been described under the assumption that resistance against CGMMV is generated "at the protein level", i.e. that the "*nucleotide sequence encoding a defective variant of the replicase gene of CGMMV*" codes for a "*defective replicase*", the expression of which at cellular level generates the desired resistance against CGMMV. However, the invention is not limited to any explanation or mechanism, and is not particularly limited to the use of a particular type of nucleotide sequence (i.e. encoding a "defective replicase"). For instance, an alternative mechanism is based on the assumption that the resistance to CGMMV is generated at the RNA-level, e.g. downregulation of gene expression due to RNA sequence homology. The use of any nucleotide sequence that can be used to provide resistance against CGMMV and that at the same time does not provide replicase activity, is envisaged by the invention.

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C L A I M S

1. Method for generating resistance in a plant or in a plant cell against infection with CGMMV, said method comprising at least the step of transforming said plant or plant cell
5 with a polynucleotide sequence that

- i) upon (at least) transformation into a plant and transcription into RNA generates resistance against infection with CGMMV in said plant; and
- ii) upon (at least) transformation into a plant and transcription into RNA does not lead to generation of (any) replicase activity in said plant.

2. Method for providing a transgenic plant and/or a transgenic plant cell that is resistant against infection with CGMMV, comprising at least the step of transforming said plant or plant cell with a polynucleotide sequence that

- i) upon (at least) transformation into a plant and transcription into RNA generates
15 resistance against infection with CGMMV in said plant; and
- ii) upon (at least) transformation into a plant and transcription into RNA does not lead to generation of (any) replicase activity in said plant.

3. Method according to claim 1 and/or 2, in which the polynucleotide sequence is a
20 nucleotide sequence that comprises, and preferably consists of:

- a nucleotide sequence corresponding to the sequence given in SEQ ID no.1, to the sequence given in SEQ ID no.17 or to the nucleotide sequence of a naturally occurring variant thereof, in which - compared to the sequence of SEQ ID no.1, SEQ ID no. 17 and/or the naturally occurring variant thereof - one or more nucleotides have been
25 added, replaced and/or removed;
- a nucleotide sequence corresponding to the sequence given in SEQ ID no.5, to the sequence given in SEQ ID no.21, to the nucleotide sequence of a naturally occurring variant thereof, in which - compared to the sequence of SEQ ID no.5, SEQ ID no. 21 and/or the naturally occurring variant thereof - one or more nucleotides have been
30 added, replaced and/or removed;
- a nucleotide sequence corresponding to the sequence given in SEQ ID no.3, to the sequence given in SEQ ID no.19 or to the nucleotide sequence of a naturally occurring variant thereof;

- a nucleotide sequence corresponding to the sequence given in SEQ ID no.3, to the sequence given in SEQ ID no.19 or to the nucleotide sequence of a naturally occurring variant thereof, in which - compared to the sequence of SEQ ID no.3, SEQ ID no. 19 and/or the naturally occurring variant thereof - one or more nucleotides have been added, replaced and/or removed;

such that said nucleotide sequence is capable, upon (at least) transformation into a plant and transcription into RNA, to confer to said plant resistance against infection with CGMMV, and such that said nucleotide sequence, upon (at least) transformation into a plant and transcription into RNA, is not capable of generating of (any) replicase activity in said plant.

4. Method according to any of claims 1-3, in which the polynucleotide sequence encodes a polypeptide or protein that

- i) upon being expressed in a plant is capable of generating resistance against CGMMV in said plant; and
- ii) upon being expressed in a plant has no replicase activity.

5. Method according to claim 4, in which the polynucleotide sequence encodes a protein or polypeptide that comprises, and preferably consists of:

- an amino acid sequence corresponding to the sequence given in SEQ ID no.2, to the sequence given in SEQ ID no.18 or to the amino acid sequence of a naturally occurring variant thereof, in which - compared to the sequence of SEQ ID no.2, SEQ ID no.18 and/or the naturally occurring variant thereof - one or more amino acids have been added, replaced or removed, preferably replaced or removed, more preferably removed;
- an amino acid sequence corresponding to the sequence given in SEQ ID no.6, to the sequence given in SEQ ID no.22 or to the amino acid sequence of a naturally occurring variant thereof, in which - compared to the sequence of SEQ ID no.6, SEQ ID no. 22 and/or the naturally occurring variant thereof - one or more amino acids have been added, replaced or removed, preferably replaced or removed, more preferably removed;
- an amino acid sequence corresponding to the sequence given in SEQ ID no.4, to the sequence given in SEQ ID no.20 or to the amino acid sequence of a naturally occurring variant thereof;

- an amino acid sequence corresponding to the sequence given in SEQ ID no.4, to the sequence given in SEQ ID no.20 or to the amino acid sequence of a naturally occurring variant thereof, in which - compared to the sequence of SEQ ID no.4, SEQ ID no. 20 and/or the naturally occurring variant thereof - one or more amino acids have been added, replaced or removed, preferably replaced or removed, more preferably removed;

or any combination thereof, provided that the resulting protein or polypeptide shows no replicase activity, but is still capable - upon expression in a plant - to generate resistance against CGMMV in said plant.

6. Method according to claim 4 or 5, in which the polynucleotide sequence encodes a protein or polypeptide that comprises, and preferably consists of :

- an amino acid sequence corresponding to a part or fragment of the sequence given in SEQ ID no.2, to a part or fragment of the sequence given in SEQ ID no.18 and/or to a part of fragment of the amino acid sequence of a naturally occurring variant thereof, or corresponding to a combination of two or more such parts or fragments;
- an amino acid sequence corresponding to a part or fragment of the sequence given in SEQ ID no.6, to a part or fragment of the sequence given in SEQ ID no.22 and/or to a part of fragment of the amino acid sequence of a naturally occurring variant thereof, or corresponding to a combination of two or more such parts or fragments; ts;
- an amino acid sequence corresponding to the sequence given in SEQ ID no.4, to the sequence given in SEQ ID no.20, and/or to the amino acid sequence of a naturally occurring variant thereof;

such that the resulting protein or polypeptide shows no replicase activity, but is still capable - upon expression in a plant - to generate resistance against CGMMV in said plant.

7. Method according to claim 4 or 5, in which the polynucleotide sequence encodes a protein or polypeptide that comprises, and preferably consists of :

- an amino acid sequence corresponding to a truncated variant of the sequence given in SEQ ID no.2, to a truncated variant of the sequence given in SEQ ID no.18 and/or to a truncated variant of the amino acid sequence of a naturally occurring variant thereof;

- an amino acid sequence corresponding to a truncated variant of the sequence given in SEQ ID no.6, to a truncated variant of the sequence given in SEQ ID no.22 and/or to a truncated variant of the amino acid sequence of a naturally occurring variant thereof;
- an amino acid sequence corresponding to the sequence given in SEQ ID no.4, to the sequence given in SEQ ID no 20, and/or to the amino acid sequence of a naturally occurring variant thereof;

such that the resulting protein or polypeptide shows no replicase activity, but is still capable - upon expression in a plant - to generate resistance against CGMMV in said plant.

8. Method according to claim 7, in which the polynucleotide sequence encodes a protein or polypeptide that comprises, and preferably consists of, the amino acid sequence given in SEQ ID no.2, the amino acid sequence given in SEQ ID no 18 and/or the amino acid sequence of a naturally occurring variant thereof, that has been truncated in the GDD-motif or in the P-loop.

9. Method according to any of the preceding claims, further comprising at least one step of cultivating the transformed plant cell into a mature plant

10. Method according to any of the preceding claims, further comprising at least one step of sexually or asexually reproducing or multiplying the transformed plant and/or the mature plant obtained from the transformed plant cell of claim 9.

11. Method according to any of the preceding claims, in which the plant is a plant that is susceptible to infection with CGMMV, more preferably a plant belonging to the *Cucurbitaceae* family, such as melon (*Cucumis melo*), cucumber (*C.sativus*), watermelon (*Citrullus vulgaris*) and bottlegourd (*Lagenaria siceraria*)

12. Genetic construct suitable for transforming a plant, said construct at least comprising nucleotide sequence that

- i) upon (at least) transformation into a plant and transcription into RNA generates resistance against infection with CGMMV in said plant; and
- ii) upon (at least) transformation into a plant and transcription into RNA does not lead to generation of (any) replicase activity in said plant;

and optionally comprising further elements of genetic constructs known per se.

13. Genetic construct according to claim 12. at least comprising a nucleotide sequence that comprises, and preferably consists of:

- 5 – a nucleotide sequence corresponding to the sequence given in SEQ ID no.1, to the sequence given in SEQ ID no.17 or to the nucleotide sequence of a naturally occurring variant thereof, in which - compared to the sequence of SEQ ID no.1, SEQ ID no. 17 and/or the naturally occurring variant thereof - one or more nucleotides have been added, replaced and/or removed;
- 10 – a nucleotide sequence corresponding to the sequence given in SEQ ID no.5, to the sequence given in SEQ ID no.21, to the nucleotide sequence of a naturally occurring variant thereof, in which - compared to the sequence of SEQ ID no.5, SEQ ID no. 21 and/or the naturally occurring variant thereof - one or more nucleotides have been added, replaced and/or removed;
- 15 – a nucleotide sequence corresponding to the sequence given in SEQ ID no.3, to the sequence given in SEQ ID no.19 or to the nucleotide sequence of a naturally occurring variant thereof;
- 20 – a nucleotide sequence corresponding to the sequence given in SEQ ID no.3, to the sequence given in SEQ ID no.19 or to the nucleotide sequence of a naturally occurring variant thereof, in which - compared to the sequence of SEQ ID no.3, SEQ ID no. 19 and/or the naturally occurring variant thereof - one or more nucleotides have been added, replaced and/or removed;

such that said construct is capable, upon (at least) transformation into a plant and transcription into RNA, to confer to said plant resistance against infection with CGMMV, and such that said construct, upon (at least) transformation into a plant and transcription into RNA, is not
25 capable of generating of (any) replicase activity in said plant.

14. Genetic construct according to claim 12 or 13, at least comprising a nucleotide sequence that encodes a polypeptide or protein that

- 30 i) upon being expressed in a plant is capable of generating resistance against CGMMV in said plant; and
- ii) upon being expressed in a plant has no replicase activity.

15. Genetic construct according to claim 14, in which the nucleotide sequence encodes a protein or polypeptide that comprises, and preferably consists of:

- 5 – an amino acid sequence corresponding to the sequence given in SEQ ID no.2, to the sequence given in SEQ ID no.18 or to the amino acid sequence of a naturally occurring variant thereof, in which - compared to the sequence of SEQ ID no.2, SEQ ID no.18 and/or the naturally occurring variant thereof - one or more amino acids have been added, replaced or removed, preferably replaced or removed, more preferably removed;
- 10 – an amino acid sequence corresponding to the sequence given in SEQ ID no.6, to the sequence given in SEQ ID no.22 or to the amino acid sequence of a naturally occurring variant thereof, in which - compared to the sequence of SEQ ID no.6, SEQ ID no. 22 and/or the naturally occurring variant thereof - one or more amino acids have been added, replaced or removed, preferably replaced or removed, more preferably removed
- 15 – an amino acid sequence corresponding to the sequence given in SEQ ID no.4, to the sequence given in SEQ ID no.20 or to the amino acid sequence of a naturally occurring variant thereof;
- 20 – an amino acid sequence corresponding to the sequence given in SEQ ID no.4, to the sequence given in SEQ ID no.20 or to the amino acid sequence of a naturally occurring variant thereof, in which - compared to the sequence of SEQ ID no.4, SEQ ID no. 20 and/or the naturally occurring variant thereof - one or more amino acids have been added, replaced or removed, preferably replaced or removed, more preferably removed;

25 or any combination thereof, provided that the resulting protein or polypeptide shows no replicase activity, but is still capable - upon expression in a plant - to generate resistance against CGMMV in said plant.

16. Genetic construct according to claim 14 or 15, in which the nucleotide sequence encodes a protein or polypeptide that comprises, and preferably consists of :

- 30 – an amino acid sequence corresponding to a part or fragment of the sequence given in SEQ ID no.2, to a part or fragment of the sequence given in SEQ ID no.18 and/or to a part of fragment of the amino acid sequence of a naturally occurring variant thereof, or corresponding to a combination of two or more such parts or fragments;

- an amino acid sequence corresponding to a part or fragment of the sequence given in SEQ ID no.6, to a part or fragment of the sequence given in SEQ ID no.22 and/or to a part of fragment of the amino acid sequence of a naturally occurring variant thereof, or corresponding to a combination of two or more such parts or fragments; ts;
- 5 – an amino acid sequence corresponding to the sequence given in SEQ ID no.4, to the sequence given in SEQ ID no.20, and/or to the amino acid sequence of a naturally occurring variant thereof;

such that the resulting protein or polypeptide shows no replicase activity, but is still capable - upon expression in a plant - to generate resistance against CGMMV in said plant.

10

17. Genetic construct according to any of claims 14-16, in which the nucleotide sequence encodes a protein or polypeptide that comprises, and preferably consists of :

- an amino acid sequence corresponding to a truncated variant of the sequence given in SEQ ID no.2, to a truncated variant of the sequence given in SEQ ID no.18 and/or to a truncated variant of the amino acid sequence of a naturally occurring variant thereof;
- 15 – an amino acid sequence corresponding to a truncated variant of the sequence given in SEQ ID no.6, to a truncated variant of the sequence given in SEQ ID no.22 and/or to a truncated variant of the amino acid sequence of a naturally occurring variant thereof;
- an amino acid sequence corresponding to the sequence given in SEQ ID no.4, to the sequence given in SEQ ID no 20, and/or to the amino acid sequence of a naturally occurring variant thereof;

20

such that the resulting protein or polypeptide shows no replicase activity, but is still capable - upon expression in a plant - to generate resistance against CGMMV in said plant.

25

18. Genetic construct according to claim 17, in which the nucleotide sequence encodes a protein or polypeptide that comprises, and preferably consists of, the amino acid sequence given in SEQ ID no.2, the amino acid sequence given in SEQ ID no.18 and/or the amino acid sequence of a naturally occurring variant thereof, that has been truncated in the GDD-motif or in the P-loop.

30

19. Genetic construct according to any of claims 12-18, in which the nucleotide sequence is under control of the plastocyanine-promoter.

20. Genetic construct according to any of claims 12-19, in which the nucleotide sequence is preceded by the native CGMMV leader (5'-UTR) sequence.

5 21. Genetic construct according to any of claims 12-20, in a form that can be stably maintained or inherited in a micro-organism, in particular a bacterium, more in particular a bacterium that can be used to transform a plant or plant material, such as *Agrobacterium*.

10 22. Micro-organism, in particular bacterium, more in particular a bacterium that can be used to transform a plant, such as *Agrobacterium*, that contains a genetic construct according to any of claims 12-21, and in particular according to claim 21.

23. Transgenic plant or plant cell, obtainable or obtained by a method according to one of claims 2-11, or a descendant of such a plant.

15 24. Plant, plant cell or plant material that has been transformed with genetic construct according to any of claims 12-21, or a descendant of such a plant.

20 25. Plant according to claim 24 or 25, being a plant that is susceptible to infection with CGMMV, more preferably a plant belonging to the *Cucurbitaceae* family, such as melon (*Cucumis melo*), cucumber (*C.sativus*), watermelon (*Citrullus vulgaris*) and bottlegourd (*Lagenaria siceraria*)

25 26. Cultivation material such as seed, tubers, roots, stalks, seedlings for a plant according to claim 23, 24 or 25.

Fig 1

6421 nucleotides

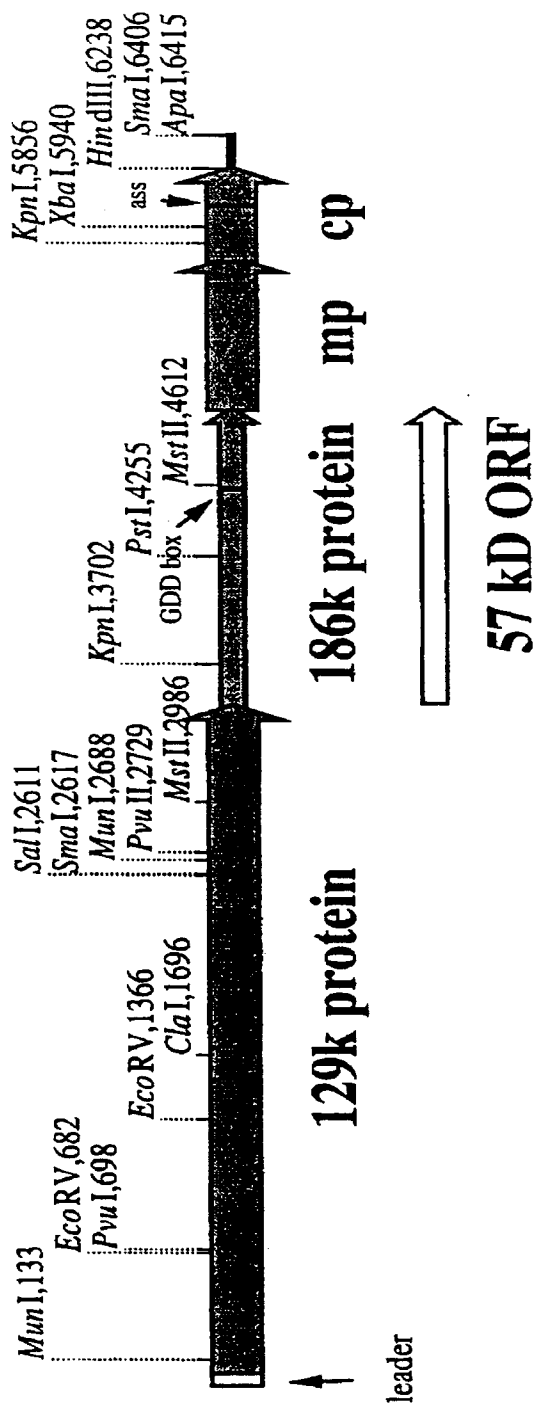


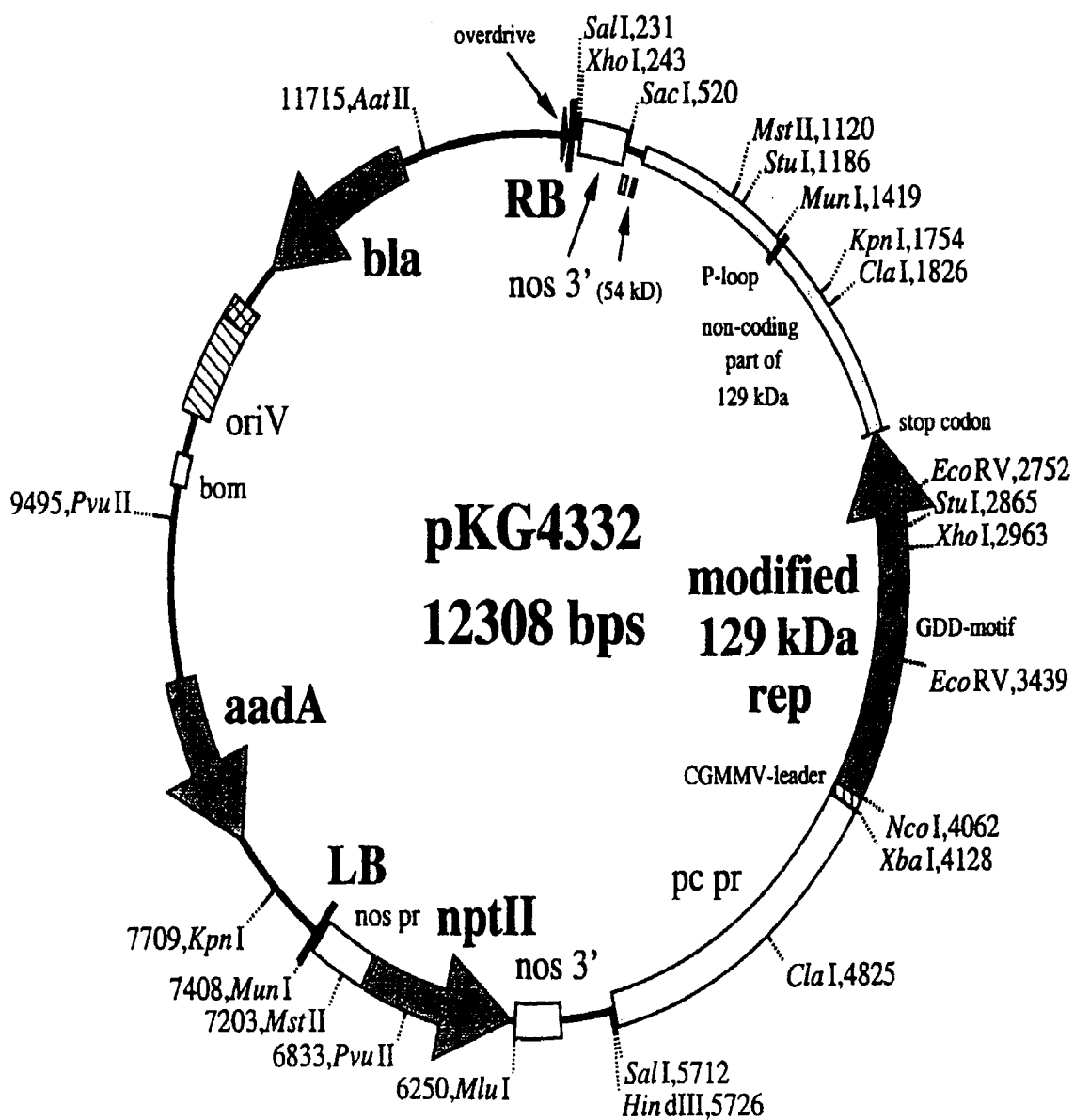
Fig 4

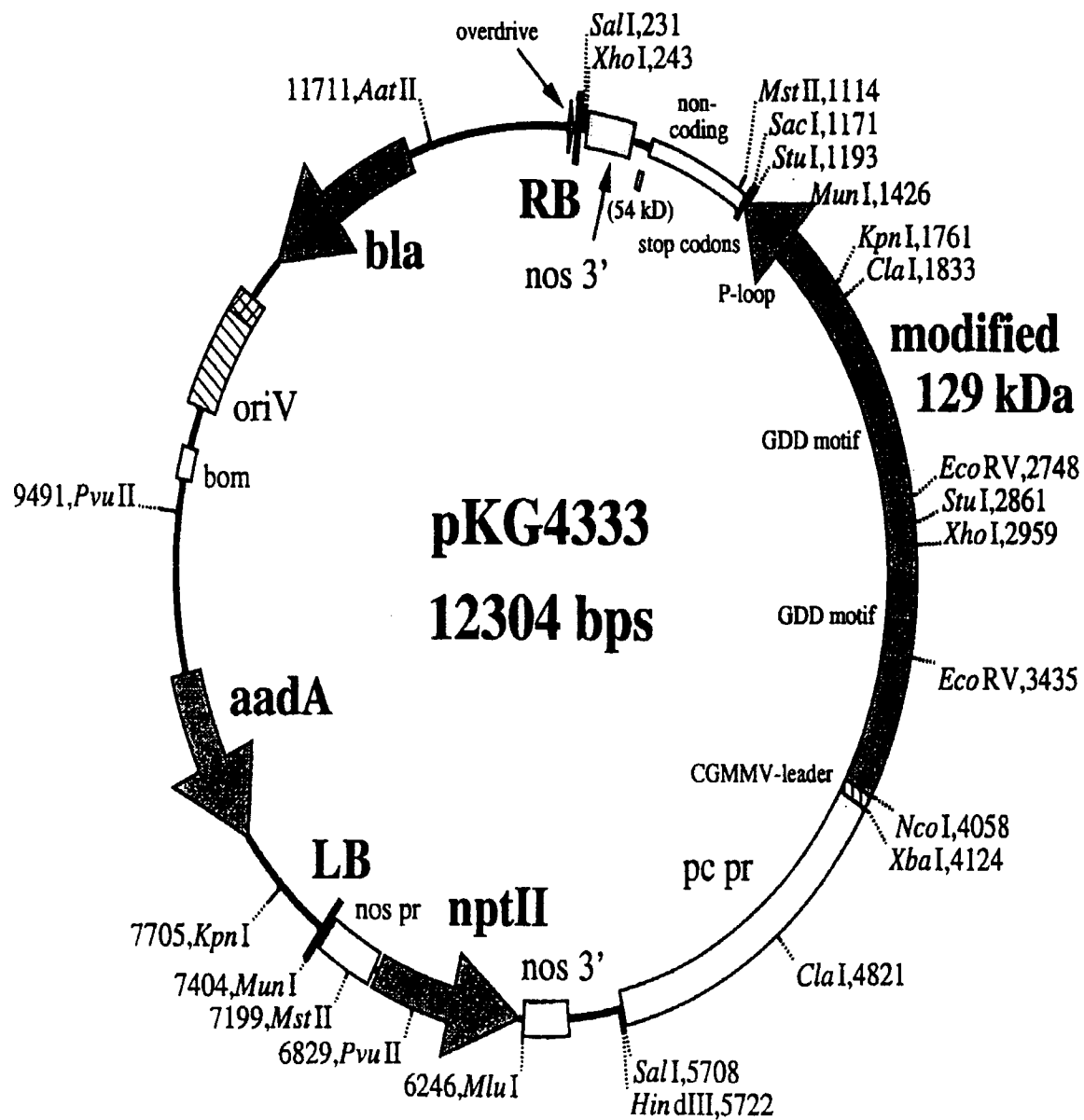
Fig 5

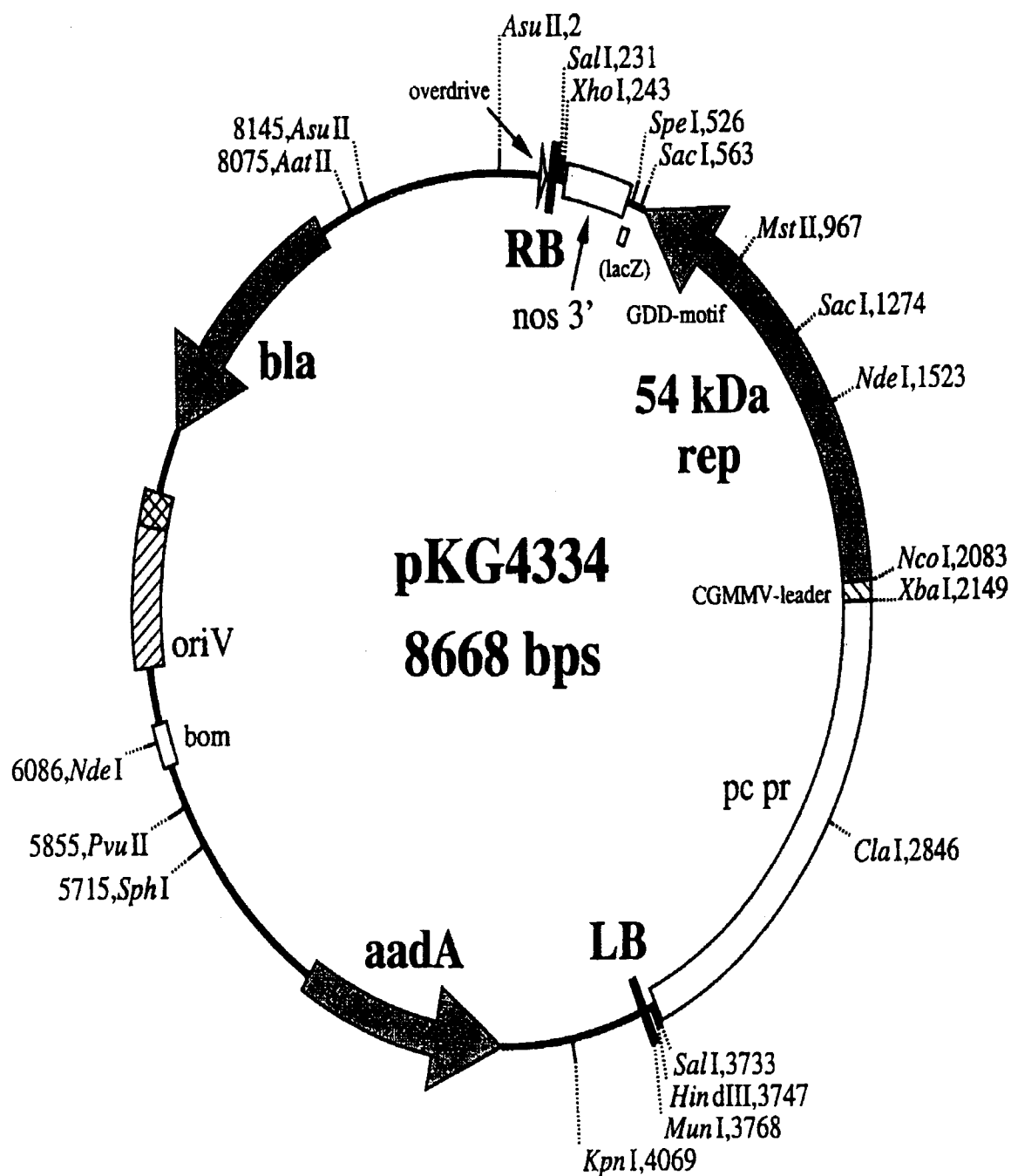
Fig 6

Fig 7

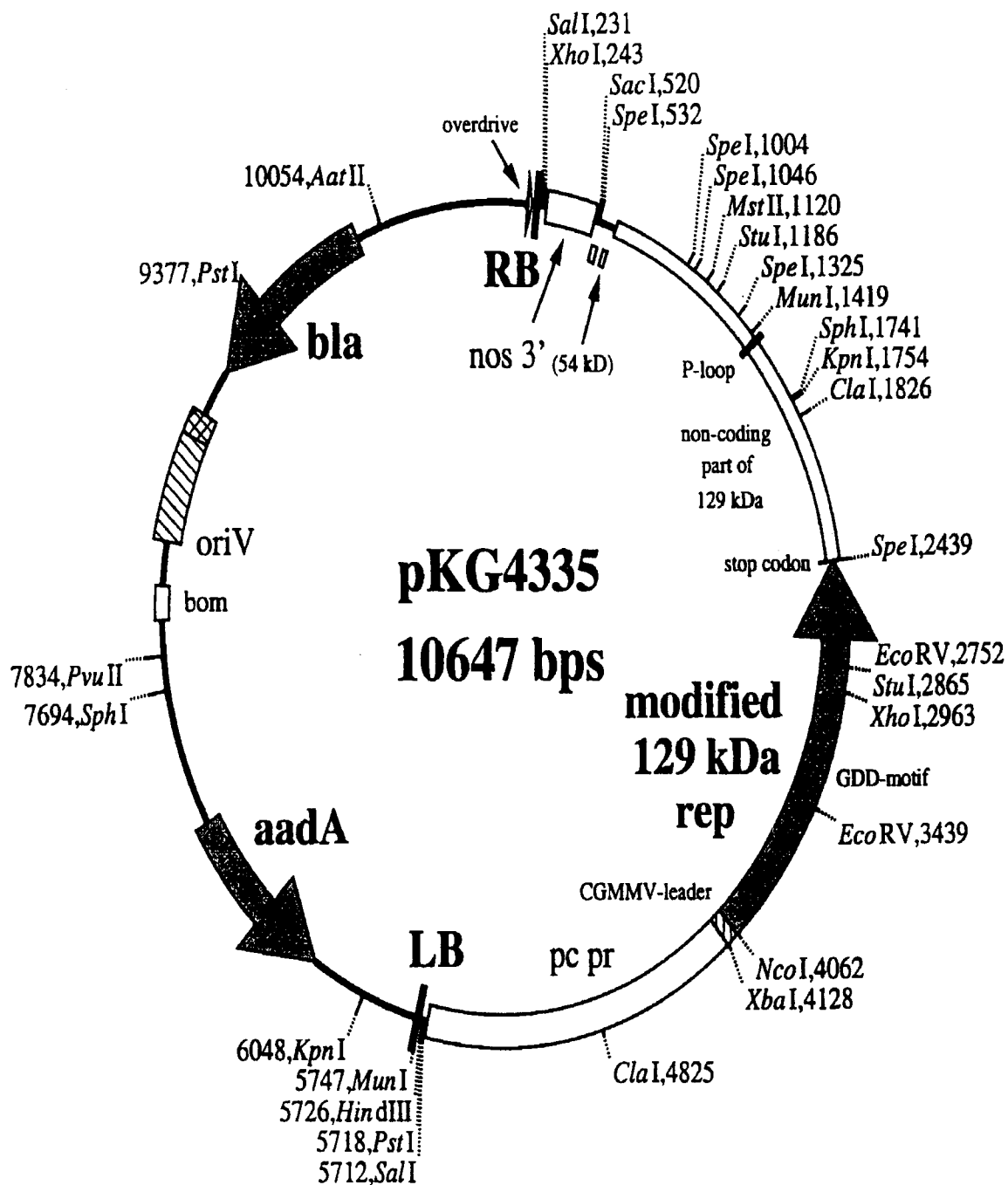
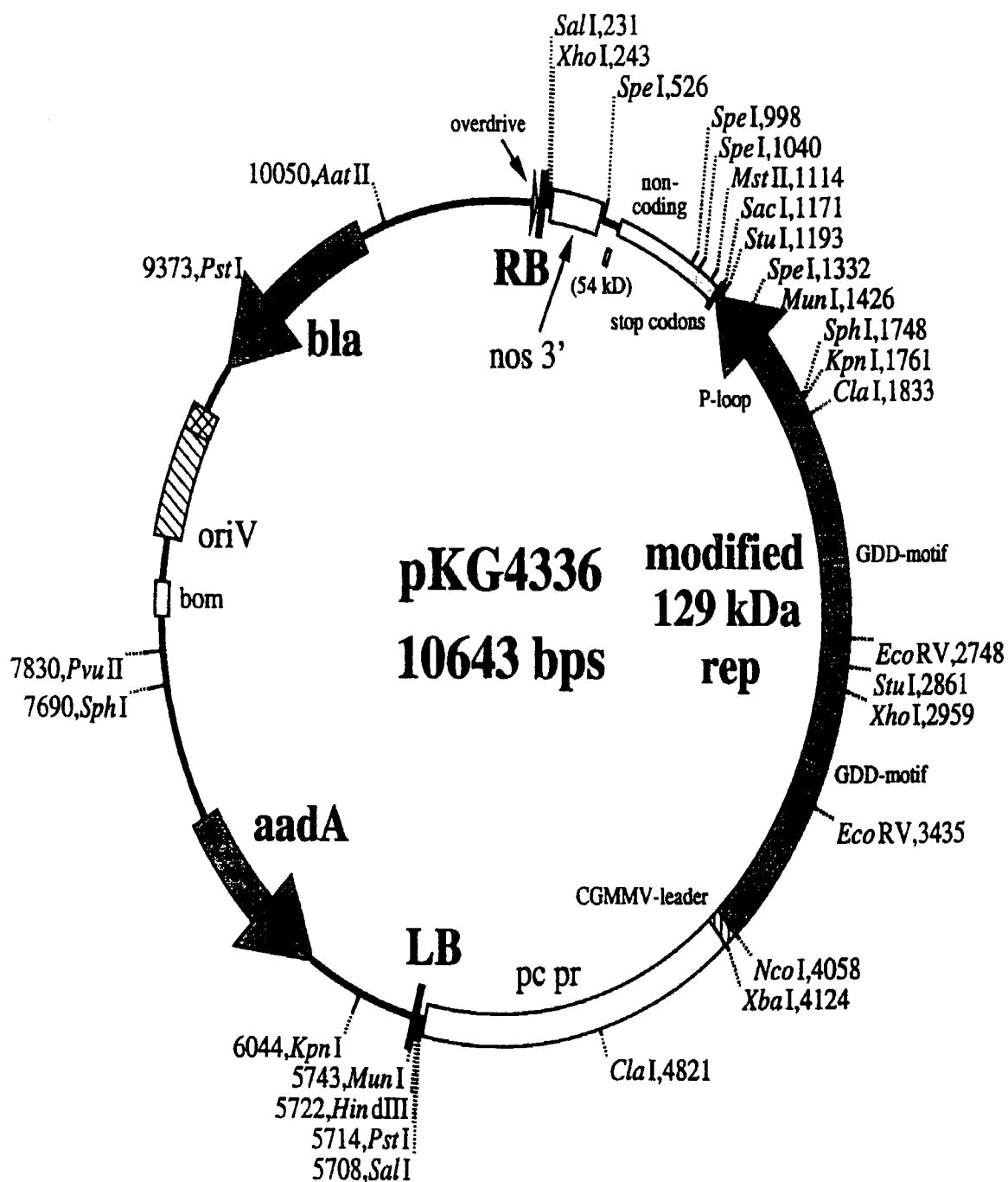


Fig 8

SEQUENCE LISTING

<110> Keygene N.V.

5 <120> Method for generating CGMMV resistance in plants,
genetic constructs for use in said method, and
CGMMV-resistant plants obtained via said method

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<170> PatentIn Ver. 2.1

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15  acgggggttg tcaactttgc gttgaagatt tcgggttgta aaaaggcctt tgtcttcggt 2880
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<210> 2

<211> 1144

30 <212> PRT

<213> Artificial Sequence

<220>

35 <223> Description of Artificial Sequence: amino acid
sequence corresponding to 129 kD replicase of
CGMMV isolate 4

<400> 2

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1 5 10 15

Ser Gly Arg Asn Asn Leu Val Ser Gln Leu Ala Ser Lys Arg Val Tyr
20 25 30

45 Asp Glu Ala Val Arg Ser Leu Asp His Gln Asp Arg Arg Pro Lys Met
35 40 45

Asn Phe Ser Arg Val Val Ser Thr Glu His Thr Arg Leu Val Thr Asp
50 55 60

Ala Tyr Pro Glu Phe Ser Ile Ser Phe Thr Ala Thr Lys Asn Ser Val
65 70 75 80

55 His Ser Leu Ala Gly Gly Leu Arg Leu Leu Glu Leu Glu Tyr Met Met
85 90 95

Met Gln Val Pro Tyr Gly Ser Pro Cys Phe Asp Ile Gly Gly Asn Tyr
100 105 110

60 Thr Gln His Leu Phe Lys Gly Arg Ser Tyr Val His Cys Cys Asn Pro
115 120 125

	Cys	Leu	Asp	Leu	Lys	Asp	Val	Ala	Arg	Asn	Val	Met	Tyr	Asn	Asp	Met	
	130						135					140					
5	Ile	Thr	Gln	His	Val	Gln	Arg	His	Lys	Gly	Ser	Gly	Gly	Cys	Arg	Pro	
	145					150					155					160	
	Leu	Pro	Thr	Phe	Gln	Ile	Asp	Ala	Phe	Arg	Arg	Tyr	Glu	Asp	Ser	Pro	
					165					170					175		
10	Val	Ala	Val	Thr	Cys	Pro	Asp	Val	Phe	Gln	Glu	Cys	Ser	Tyr	Asp	Phe	
				180					185						190		
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15			195					200					205				
	Ile	Pro	Tyr	Ser	Ser	Ile	Gly	Pro	Ala	Leu	His	Arg	Lys	Asn	Val	Arg	
	210						215					220					
20	Val	Cys	Tyr	Ala	Ala	Phe	His	Phe	Ser	Glu	Ala	Leu	Leu	Leu	Gly	Ser	
	225					230					235					240	
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					245					250					255		
25	Asp	Asp	Val	His	Phe	Leu	Phe	Ser	Glu	Glu	Ser	Thr	Leu	His	Tyr	Thr	
				260					265						270		
	His	Ser	Leu	Glu	Asn	Ile	Lys	Leu	Ile	Val	Met	Arg	Thr	Tyr	Phe	Pro	
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	290						295					300					
35	Asp	Thr	Phe	Phe	Phe	Arg	Leu	Val	Arg	Ala	Asp	Thr	His	Met	Leu	His	
	305					310					315					320	
	Lys	Ser	Val	Gly	His	Tyr	Ser	Lys	Ser	Lys	Ser	Glu	Tyr	Phe	Ala	Leu	
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40	Asn	Thr	Pro	Pro	Ile	Phe	Gln	Asp	Lys	Ala	Thr	Phe	Ser	Val	Trp	Phe	
			340						345					350			
	Pro	Glu	Ala	Lys	Arg	Lys	Val	Leu	Ile	Pro	Lys	Phe	Glu	Leu	Ser	Arg	
45			355					360					365				
	Phe	Leu	Ser	Gly	Asn	Val	Lys	Val	Ser	Arg	Met	Leu	Val	Asp	Ala	Asp	
	370						375					380					
50	Phe	Val	His	Thr	Ile	Ile	Asn	His	Ile	Ser	Thr	Tyr	Asp	Asn	Lys	Ala	
	385				390						395					400	
	Leu	Val	Trp	Lys	Asn	Val	Gln	Ser	Phe	Val	Glu	Ser	Ile	Arg	Ser	Arg	
				405						410					415		
55	Val	Ile	Val	Asn	Gly	Val	Ser	Val	Lys	Ser	Glu	Trp	Asn	Val	Pro	Val	
				420					425					430			
	Asp	Gln	Leu	Thr	Asp	Ile	Ser	Phe	Ser	Ile	Phe	Leu	Leu	Val	Lys	Val	
60			435					440					445				

	Arg	Lys	Val	Gln	Ile	Glu	Leu	Met	Ser	Asp	Lys	Val	Val	Ile	Glu	Ala	
	450						455					460					
5	Arg	Gly	Leu	Leu	Arg	Arg	Phe	Ala	Asp	Ser	Leu	Lys	Ser	Ala	Val	Glu	
	465					470					475					480	
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					485					490					495		
10	Asp	Thr	Ser	Ser	Asp	Glu	Leu	Lys	Val	Leu	Leu	Pro	Glu	Pro	Phe	Met	
				500					505					510			
	Thr	Phe	Ser	Asp	Tyr	Leu	Glu	Gly	Met	Tyr	Glu	Ala	Asp	Ala	Lys	Ile	
			515					520					525				
15	Glu	Arg	Glu	Ser	Val	Ser	Glu	Leu	Leu	Ala	Ser	Gly	Asp	Asp	Leu	Phe	
		530					535					540					
	Lys	Lys	Ile	Asp	Glu	Ile	Arg	Asn	Asn	Tyr	Ser	Gly	Val	Glu	Phe	Asp	
20		545				550					555					560	
	Val	Glu	Lys	Phe	Gln	Glu	Phe	Cys	Lys	Glu	Leu	Asn	Val	Asn	Pro	Met	
				565						570					575		
25	Leu	Ile	Gly	His	Val	Ile	Glu	Ala	Ile	Phe	Ser	Gln	Lys	Ala	Gly	Val	
				580					585					590			
	Thr	Val	Thr	Gly	Leu	Gly	Thr	Leu	Ser	Pro	Glu	Met	Gly	Ala	Ser	Val	
			595					600					605				
30	Ala	Leu	Ser	Asn	Asn	Ser	Val	Asp	Thr	Cys	Asp	Asp	Met	Asp	Val	Thr	
		610					615					620					
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				645					650						655		
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			660						665					670			
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			675					680					685				
45	Cys	Val	Arg	Lys	Pro	Gln	Phe	Ser	Lys	Pro	Leu	Asp	Glu	Glu	Asp	Asp	
		690					695					700					
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	Lys	Lys	Thr	Ile	Thr	Pro	Val	Val	Tyr	Thr	Gly	Thr	Ile	Arg	Glu	Arg	
				725						730					735		
55	Gln	Met	Lys	Asn	Tyr	Ile	Asp	Tyr	Leu	Ser	Ala	Ser	Leu	Gly	Ser	Thr	
				740					745					750			
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			755				760						765				
60	Glu	Ser	Met	Gln	Thr	Phe	Gly	Leu	Tyr	Asp	Cys	Glu	Lys	Cys	Lys	Trp	

	770		775		780
	Leu Leu Leu Pro Ser Glu Lys Lys His Ala Trp Ala Val Val Leu Ala				
	785		790		795 800
5	Ser Asp Asp Thr Thr Arg Ile Ile Phe Leu Ser Tyr Asp Glu Ser Gly				
		805		810	815
	Ser Pro Ile Ile Asp Lys Lys Asn Trp Lys Arg Phe Ala Val Cys Ser				
10		820		825	830
	Asp Thr Lys Val Tyr Ser Val Ile Arg Ser Leu Glu Val Leu Asn Lys				
		835		840	845
15	Glu Ala Thr Val Asp Pro Gly Val Tyr Ile Thr Leu Val Asp Gly Val				
	850		855		860
	Pro Gly Cys Gly Lys Thr Ala Glu Ile Ile Ala Arg Val Asn Trp Lys				
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	Thr Asp Leu Val Leu Thr Pro Gly Arg Glu Ala Ala Ala Met Ile Arg				
		885		890	895
	Arg Arg Ala Cys Ala Leu His Lys Ser Pro Val Ala Thr Ser Asp Asn				
25		900		905	910
	Val Arg Thr Phe Asp Ser Phe Val Met Asn Lys Lys Val Phe Lys Phe				
		915		920	925
30	Asp Ala Val Tyr Val Asp Glu Gly Leu Met Val His Thr Gly Leu Leu				
	930		935		940
	Asn Phe Ala Leu Lys Ile Ser Gly Cys Lys Lys Ala Phe Val Phe Gly				
35	945		950		955 960
	Asp Ala Lys Gln Ile Pro Phe Ile Asn Arg Val Met Asn Phe Asp Tyr				
		965		970	975
	Pro Lys Glu Leu Arg Thr Leu Ile Val Asp Asn Val Glu Arg Arg Tyr				
40		980		985	990
	Ile Thr His Arg Cys Pro Arg Asp Val Thr Ser Phe Leu Asn Thr Ile				
		995		1000	1005
45	Tyr Lys Ala Ala Val Ser Thr Thr Ser Pro Val Val His Ser Val Lys				
	1010		1015		1020
	Ala Ile Lys Val Ser Gly Ala Gly Ile Leu Arg Pro Glu Leu Thr Lys				
50	1025		1030		1035 1040
	Ile Lys Gly Lys Ile Ile Thr Phe Thr Gln Ser Asp Lys Gln Ser Leu				
		1045		1050	1055
	Ile Lys Ser Gly Tyr Asn Asp Val Asn Thr Val His Glu Ile Gln Gly				
55		1060		1065	1070
	Glu Thr Phe Glu Glu Thr Ala Val Val Arg Ala Thr Pro Thr Pro Ile				
		1075		1080	1085
60	Gly Leu Ile Ala Arg Asp Ser Pro His Val Leu Val Ala Leu Thr Arg				
	1090		1095		1100

His Thr Lys Ala Met Val Tyr Tyr Thr Val Val Phe Asp Ala Val Thr
 1105 1110 1115 1120

5 Ser Ile Ile Ala Asp Val Glu Lys Val Asp Gln Ser Ile Leu Thr Met
 1125 1130 1135

Phe Ala Thr Thr Val Pro Thr Lys
 1140

10

<210> 3

<211> 1503

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<212> DNA

<213> Artificial Sequence

<220>

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<223> Description of Artificial Sequence: nucleotide
 sequence corresponding to 57 kD protein of CGMMV
 isolate 4

<400> 3

25

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gatttcgatg ccgtcaccat gcggttgagg gataatgaat tcaatttgca accttgtaga 180
ttaactttaa gtaatttaga tccggtgccg gctttgatta agagtgaggc aaaagatttt 240
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30 actaatatgt ctatttctat agtagataat ttcttttctt cctttgtcag ggacgagggt 420
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50

<210> 4

<211> 501

<212> PRT

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<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: amino acid
 sequence corresponding to 57 kD protein of CGMMV
 isolate 4

60

<400> 4

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				20					25					30			
	Pro	Gly	Asn	Ser	Phe	Val	Leu	Asn	Asp	Phe	Asp	Ala	Val	Thr	Met	Arg	
			35					40					45				
10	Leu	Arg	Asp	Asn	Glu	Phe	Asn	Leu	Gln	Pro	Cys	Arg	Leu	Thr	Leu	Ser	
	50						55					60					
	Asn	Leu	Asp	Pro	Val	Pro	Ala	Leu	Ile	Lys	Ser	Glu	Ala	Lys	Asp	Phe	
15	65					70				75						80	
	Leu	Val	Pro	Val	Leu	Arg	Thr	Ala	Cys	Glu	Arg	Pro	Arg	Ile	Pro	Gly	
					85					90					95		
20	Leu	Leu	Glu	Asn	Leu	Val	Ala	Met	Ile	Lys	Arg	Asn	Met	Asn	Thr	Pro	
				100					105					110			
	Asp	Leu	Ala	Gly	Thr	Val	Asp	Ile	Thr	Asn	Met	Ser	Ile	Ser	Ile	Val	
			115					120					125				
25	Asp	Asn	Phe	Phe	Ser	Ser	Phe	Val	Arg	Asp	Glu	Val	Leu	Leu	Asp	His	
			130				135					140					
	Leu	Asp	Cys	Val	Arg	Ala	Ser	Ser	Ile	Gln	Ser	Phe	Ser	Asp	Trp	Phe	
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35	Ile	Asp	Leu	Pro	Ala	Phe	Asp	Thr	Tyr	Met	His	Met	Ile	Lys	Arg	Gln	
				180					185					190			
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					245					250					255		
50	Phe	Ser	Asp	Leu	Ser	Ser	His	Ser	Asp	Tyr	Glu	Ile	Leu	Glu	Leu	Asp	
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	Val	Ser	Lys	Tyr	Asp	Lys	Ser	Gln	Ser	Asp	Phe	His	Phe	Ser	Ile	Glu	
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	Trp	Ser	Met	Gly	His	Lys	Arg	Thr	Ile	Leu	Gln	Asp	Phe	Gln	Ala	Gly	
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Ile Lys Thr Leu Ile Tyr Tyr Gln Arg Lys Ser Gly Asp Val Thr Thr
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5 Phe Ile Gly Asn Thr Phe Ile Ile Ala Ala Cys Val Ala Ser Met Leu
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Pro Leu Asp Lys Cys Phe Lys Ala Ser Phe Cys Gly Asp Asp Ser Leu
 355 360 365

10 Ile Tyr Leu Pro Lys Gly Leu Glu Tyr Pro Asp Ile Gln Ala Thr Ala
 370 375 380

Asn Leu Val Trp Asn Phe Glu Ala Lys Leu Phe Arg Lys Lys Tyr Gly
 385 390 395 400

15 Tyr Phe Cys Gly Lys Tyr Ile Ile His His Ala Asn Gly Cys Ile Val
 405 410 415

20 Tyr Pro Asp Pro Leu Lys Leu Ile Ser Lys Leu Gly Ser Lys Ser Leu
 420 425 430

Val Gly Tyr Glu His Val Glu Glu Phe Arg Ile Ser Leu Leu Asp Val
 435 440 445

25 Ala His Ser Leu Phe Asn Gly Ala Tyr Phe His Leu Leu Asp Asp Ala
 450 455 460

Ile His Glu Leu Phe Pro Asn Ala Gly Gly Cys Ser Phe Val Ile Asn
 465 470 475 480

30 Cys Leu Cys Lys Tyr Leu Ser Asp Lys Arg Leu Phe Arg Ser Leu Tyr
 485 490 495

35 Ile Asp Val Ser Lys
 500

<210> 5
 40 <211> 4935
 <212> DNA
 <213> Artificial Sequence

<220>
 45 <223> Description of Artificial Sequence: nucleotide
 sequence corresponding to 186 kD protein of CGMMV
 isolate 4

<400> 5
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           20             25             30

30 Asp Glu Ala Val Arg Ser Leu Asp His Gln Asp Arg Arg Pro Lys Met
      35             40             45

    Asn Phe Ser Arg Val Val Ser Thr Glu His Thr Arg Leu Val Thr Asp
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35 Ala Tyr Pro Glu Phe Ser Ile Ser Phe Thr Ala Thr Lys Asn Ser Val
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    Met Gln Val Pro Tyr Gly Ser Pro Cys Phe Asp Ile Gly Gly Asn Tyr
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 gaatcgctta acgcagttaa gcgtactgac gatgcgtcta cagccgctag ggctgagata 900
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 tctgacgggtg gtgcacacca tagtgcatag tgctttcccg ttcactttaa tcgaacgggt 1080
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<210> 9
 <211> 1139
 <212> DNA
 <213> Artificial Sequence

35

<220>
 <223> Description of Artificial Sequence: nucleotide
 sequence corresponding to sequence encoding coat
 protein of CGMMV isolate 3

40

<400> 9
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 gggtcatccc taactattct gtcgtggttg cggatgccct tcgcgatcct tggctcttat 180
 ttgtgaggct ctctaacgta ggtattaagg atggttttca tccattaact ttagaggctg 240
 cctgtctagt tgccactact aactctatta ttaaaaagg gcttagagct tctgtagttg 300
 agtccgttgt ctcttcogat cagtcgattg ttctagattc tttatctgag aaagttgagc 360
 50 ctttcttcga taaagtccct atttcagcgg ctgtaatggc gagagacccc agttataggt 420
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 55 gagtctttgt ctgcgttacc ttcacccgtc gtagatatta attctaggtt cccgagtgcg 720
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<210> 10

5 <211> 1139

<212> DNA

<213> Artificial Sequence

<220>

10 <223> Description of Artificial Sequence: nucleotide
sequence corresponding to sequence encoding coat
protein of CGMMV isolate 4

<400> 10

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20 agtccgttgt ctcttcogat cagtcgattg ttctagattc tttatctgag aaagttgagc 360
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35

<210> 11

<211> 1139

<212> DNA

<213> Artificial Sequence

40

<220>

<223> Description of Artificial Sequence: nucleotide
sequence corresponding to sequence encoding coat
protein of CGMMV isolate 5

45

<400> 11

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5

<210> 12

<211> 1139

<212> DNA

10 <213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: nucleotide
 sequence corresponding to sequence encoding coat
 protein of CGMMV isolate 6

15

<400> 12

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 gggtcatccc taactattct gtcgtggttg cggatgccct tcgcgatacct tgggtctttat 180
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 ctttcttcga taaagtcctt atttcagcgg ctgtaatggc gagagacccc agttataggt 420
 25 ctaggtcgca gtctgtcgtt ggtcgtggta agcggcattc taaacctcca aatcggaggt 480
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 35 tctgacgggtg gtgcacacca tagtgcatag tgctttcccg ttcactttta tcgaacgggt 1080
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<210> 13

40 <211> 1138

<212> DNA

<213> Artificial Sequence

<220>

45 <223> Description of Artificial Sequence: nucleotide
 sequence corresponding to sequence encoding coat
 protein of CGMMV isolate 7

<400> 13

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 55 agtccggttg ctcttccgat cagtcgattg ttctagattc tttatctgag aaagttgagc 360
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 60 tttctagtgg cgtcgcaagg tactgctttc caaaccagcagg caggaagaga ttccttccgt 660
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ggttttttacg ctttcctcaa cggctcctgtg ttgaggccta tcttcgtttc gcttcttagc 780
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10 <210> 14
    <211> 1139
    <212> DNA
    <213> Artificial Sequence

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15 <220>
    <223> Description of Artificial Sequence: nucleotide
          sequence corresponding to sequence encoding coat
          protein of CGMMV isolate 8

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ggttcatccc taactattct gtcgtgggtg cggatgccct tcgcgaccc ttgtctttat 180
ttgtgaggct ctctaacgta ggtattaagg atggttttca tccattaact ttagaggctg 240
25 cctgtctagt tgccactact aactctatta ttaaaaaggg gcttagagct tctgtagtgt 300
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40
    <210> 15
    <211> 1139
    <212> DNA
    <213> Artificial Sequence

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    <220>
    <223> Description of Artificial Sequence: nucleotide
          sequence corresponding to sequence encoding coat
          protein of CGMMV isolate 9

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55 ggttcatccc taactattct gtcgtgggtg cggatgccct tcgcgaccc ttgtctttat 180
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60 ctaggtcaca gtctgtcgtt ggtcgtggta agcggcattc taaacctcca aatcggaggt 480
cggactctgc ttctgaagag tccagttctg tttcttttga agatggctta caatccgac 540

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acgcctagca aacttattgc gtttagtgct tcatatgttc ccgttagaac tttacttaat 600
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<210> 16

<211> 1139

15 <212> DNA

<213> Artificial Sequence

<220>

20 <223> Description of Artificial Sequence: nucleotide
sequence corresponding to sequence encoding coat
protein of CGMMV isolate 10.

<400> 16

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25 ttgcagaagg aactatatgc aaattttcag ctcccgcac cgccgcgag ttctctgtta 120
ggttcattcc taactattct gtcgtggctg cggatgccct tcgcgaccc tggcttttat 180
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45 <210> 17

<211> 3429

<212> DNA

<213> Artificial Sequence

50 <220>

<223> Description of Artificial Sequence: nucleotide
sequence corresponding to 129 kD replicase of
CGMMV strain SH

55 <400> 17

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catcaagaca gacgcccga aatgaatttt tctcgtgtgg tcagcacaga gcacaccagg 180
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45 actagttttt ttaatactat ctataaagcc gctgtcgcta ctactagtcc ggttgtagat 3060
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cgtgccaccc cgactccaat aggtttgatt gcccggtgat caccacatgt actagtggcc 3300
50 ttaactaggc aactaaggc aatggtgtat tatactgttg tattcgatgc agttacaagt 3360
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```

55 <210> 18
 <211> 1143
 <212> PRT
 <213> Artificial Sequence

60 <220>
 <223> Description of Artificial Sequence: amino acid

sequence corresponding to 129 kD replicase of
CGMMV strain SH

<400> 18

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	Ser	Gly	Arg	Asn	Asn	Leu	Val	Ser	Gln	Leu	Ala	Ser	Lys	Arg	Val	Tyr	20	25	30	
10	Asp	Glu	Ala	Val	Arg	Ser	Leu	Asp	His	Gln	Asp	Arg	Arg	Pro	Lys	Met	35	40	45	
	Asn	Phe	Ser	Arg	Val	Val	Ser	Thr	Glu	His	Thr	Arg	Leu	Val	Thr	Asp	50	55	60	
15	Ala	Tyr	Pro	Glu	Phe	Ser	Ile	Ser	Phe	Thr	Ala	Thr	Lys	Asn	Ser	Val	65	70	75	80
20	His	Ser	Leu	Ala	Gly	Gly	Leu	Arg	Leu	Leu	Glu	Leu	Glu	Tyr	Met	Met	85	90	95	
	Met	Gln	Val	Pro	Tyr	Gly	Ser	Pro	Cys	Tyr	Asp	Ile	Gly	Gly	Asn	Tyr	100	105	110	
25	Thr	Gln	His	Leu	Phe	Lys	Gly	Arg	Ser	Tyr	Val	His	Cys	Cys	Asn	Pro	115	120	125	
30	Cys	Leu	Asp	Leu	Lys	Asp	Val	Ala	Arg	Asn	Val	Met	Tyr	Asn	Asp	Met	130	135	140	
	Ile	Thr	Gln	His	Val	Gln	Arg	His	Lys	Gly	Ser	Cys	Gly	Cys	Arg	Pro	145	150	155	160
35	Leu	Pro	Thr	Phe	Gln	Ile	Asp	Ala	Phe	Arg	Arg	Tyr	Asp	Ser	Ser	Pro	165	170	175	
	Cys	Ala	Val	Thr	Cys	Ser	Asp	Val	Phe	Gln	Glu	Cys	Ser	Tyr	Asp	Phe	180	185	190	
40	Gly	Ser	Gly	Arg	Asp	Asn	His	Ala	Val	Ser	Leu	His	Ser	Ile	Tyr	Asp	195	200	205	
45	Ile	Pro	Tyr	Ser	Ser	Ile	Gly	Pro	Ala	Leu	His	Arg	Lys	Asn	Val	Arg	210	215	220	
	Val	Cys	Tyr	Ala	Ala	Phe	His	Phe	Ser	Glu	Ala	Leu	Leu	Leu	Gly	Ser	225	230	235	240
50	Pro	Val	Gly	Asn	Leu	Asn	Ser	Ile	Gly	Ala	Gln	Phe	Arg	Val	Asp	Gly	245	250	255	
	Asp	Asp	Val	His	Phe	Leu	Phe	Ser	Glu	Glu	Ser	Thr	Leu	His	Tyr	Thr	260	265	270	
55	His	Ser	Leu	Glu	Asn	Ile	Lys	Leu	Ile	Val	Met	Arg	Thr	Tyr	Phe	Pro	275	280	285	
60	Ala	Asp	Asp	Arg	Phe	Val	Tyr	Ile	Lys	Glu	Phe	Met	Val	Lys	Arg	Val	290	295	300	

	Asp	Thr	Phe	Phe	Phe	Arg	Leu	Val	Arg	Ala	Asp	Thr	His	Met	Leu	His	305	310	315	320
5	Lys	Ser	Val	Gly	His	Tyr	Ser	Lys	Ser	Lys	Ser	Glu	Tyr	Phe	Ala	Leu		325	330	335
	Asn	Thr	Pro	Pro	Ile	Phe	Gln	Asp	Lys	Ala	Thr	Phe	Ser	Val	Trp	Phe		340	345	350
10	Pro	Glu	Ala	Lys	Lys	Val	Leu	Ile	Pro	Lys	Phe	Glu	Leu	Ser	Arg	Phe		355	360	365
	Leu	Ser	Gly	Asn	Val	Lys	Ile	Ser	Arg	Met	Leu	Val	Asp	Ala	Asp	Phe		370	375	380
15	Val	His	Thr	Ile	Ile	Asn	His	Ile	Ser	Thr	Tyr	Asp	Asn	Lys	Ala	Leu		385	390	400
	Val	Trp	Lys	Asn	Val	Gln	Ser	Phe	Val	Glu	Ser	Ile	Arg	Ser	Arg	Val		405	410	415
20	Ile	Val	Asn	Gly	Val	Ser	Val	Lys	Ser	Glu	Trp	Asn	Val	Pro	Val	Asp		420	425	430
	Gln	Leu	Thr	Asp	Ile	Ser	Phe	Ser	Ile	Phe	Pro	Leu	Val	Lys	Val	Arg		435	440	445
	Lys	Val	Gln	Ile	Glu	Leu	Met	Ser	Asp	Lys	Val	Val	Ile	Glu	Ala	Arg		450	455	460
30	Gly	Leu	Leu	Arg	Arg	Phe	Ala	Asp	Ser	Leu	Lys	Ser	Ala	Val	Glu	Gly		465	470	480
	Leu	Gly	Asp	Cys	Val	Tyr	Asp	Ala	Leu	Val	Gln	Thr	Gly	Trp	Phe	Asp		485	490	495
35	Thr	Ser	Ser	Asp	Glu	Leu	Lys	Val	Leu	Leu	Pro	Glu	Pro	Phe	Met	Thr		500	505	510
	Phe	Ser	Asp	Tyr	Leu	Glu	Gly	Met	Tyr	Glu	Ala	Asp	Ala	Lys	Ile	Glu		515	520	525
	Arg	Glu	Ser	Val	Ser	Glu	Leu	Leu	Ala	Ser	Gly	Asp	Asp	Leu	Phe	Lys		530	535	540
45	Lys	Ile	Asp	Glu	Ile	Arg	Asn	Asn	Tyr	Ser	Gly	Val	Glu	Phe	Asp	Val		545	550	555
	Glu	Lys	Phe	Gln	Glu	Phe	Cys	Lys	Glu	Leu	Asn	Val	Asn	Pro	Met	Leu		565	570	575
50	Ile	Gly	His	Val	Ile	Glu	Ala	Ile	Phe	Ser	Gln	Lys	Ala	Gly	Val	Thr		580	585	590
	Val	Thr	Gly	Leu	Gly	Thr	Leu	Ser	Pro	Glu	Met	Gly	Ala	Ser	Val	Ala		595	600	605
	Leu	Ser	Ser	Thr	Ser	Val	Asp	Thr	Cys	Glu	Asp	Met	Asp	Val	Thr	Glu		610	615	620
60	Asp	Met	Glu	Asp	Ile	Val	Leu	Met	Ala	Asp	Lys	Ser	His	Ser	Tyr	Met				

	625				630				635				640				
5	Ser	Pro	Glu	Met	Ala	Arg	Trp	Ala	Asp	Val	Lys	Tyr	Gly	Asn	Asn	Lys	
					645					650	655						
	Gly	Ala	Leu	Val	Glu	Tyr	Lys	Val	Gly	Thr	Ser	Met	Thr	Leu	Pro	Ala	
					660					665	670						
10	Thr	Trp	Ala	Glu	Lys	Val	Lys	Ala	Val	Leu	Pro	Leu	Ser	Gly	Ile	Cys	
					675	680				685							
	Val	Arg	Lys	Pro	Gln	Phe	Ser	Lys	Pro	Leu	Asp	Glu	Glu	Asp	Asp	Leu	
					690	695				700							
15	Arg	Leu	Ser	Asn	Met	Asn	Phe	Phe	Lys	Val	Ser	Asp	Leu	Lys	Leu	Lys	
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	Lys	Thr	Ile	Thr	Pro	Val	Val	Tyr	Thr	Gly	Thr	Ile	Arg	Glu	Arg	Gln	
20					725					730				735			
	Met	Lys	Asn	Tyr	Ile	Asp	Tyr	Leu	Ser	Ala	Ser	Leu	Gly	Ser	Thr	Leu	
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25	Gly	Asn	Leu	Glu	Arg	Ile	Val	Arg	Ser	Asp	Trp	Asn	Gly	Thr	Glu	Glu	
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	Ser	Met	Gln	Thr	Phe	Gly	Leu	Tyr	Asp	Cys	Glu	Lys	Cys	Lys	Trp	Leu	
					770	775				780							
30	Leu	Leu	Pro	Ala	Glu	Lys	Lys	His	Ala	Trp	Ala	Val	Val	Leu	Ala	Ser	
					785	790				795				800			
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35					805					810				815			
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40	Thr	Lys	Val	Tyr	Ser	Val	Ile	Arg	Ser	Leu	Glu	Val	Leu	Asn	Lys	Glu	
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45	Gly	Cys	Gly	Lys	Thr	Ala	Glu	Ile	Ile	Ala	Arg	Val	Asn	Trp	Lys	Thr	
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	Asp	Leu	Val	Leu	Thr	Pro	Gly	Arg	Glu	Ala	Ala	Ala	Met	Ile	Arg	Arg	
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	Arg	Ala	Cys	Ala	Leu	His	Lys	Ser	Pro	Val	Ala	Thr	Asn	Asp	Asn	Val	
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55	Arg	Thr	Phe	Asp	Ser	Phe	Val	Met	Asn	Arg	Lys	Ile	Phe	Lys	Phe	Asp	
					915	920				925							
	Ala	Val	Tyr	Val	Asp	Glu	Gly	Leu	Met	Val	His	Thr	Gly	Leu	Leu	Asn	
					930	935				940							
60	Phe	Ala	Leu	Lys	Ile	Ser	Gly	Cys	Lys	Lys	Ala	Phe	Val	Phe	Gly	Asp	
					945	950				955				960			

Ala Lys Gln Ile Pro Phe Ile Asn Arg Val Met Asn Phe Asp Tyr Pro
 965 970 975

5 Lys Glu Leu Arg Thr Leu Ile Val Asp Asn Val Glu Arg Arg Tyr Val
 980 985 990

Thr His Arg Cys Pro Arg Asp Val Thr Ser Phe Leu Asn Thr Ile Tyr
 995 1000 1005

10 Lys Ala Ala Val Ala Thr Thr Ser Pro Val Val His Ser Val Lys Ala
 1010 1015 1020

Ile Lys Val Ser Gly Ala Gly Ile Leu Arg Pro Glu Leu Thr Lys Ile
 15 1025 1030 1035 1040

Lys Gly Lys Ile Ile Thr Phe Thr Gln Ser Asp Lys Gln Ser Leu Ile
 1045 1050 1055

20 Lys Ser Gly Tyr Asn Asp Val Asn Thr Val His Glu Ile Gln Gly Glu
 1060 1065 1070

Thr Phe Glu Glu Thr Ala Val Val Arg Ala Thr Pro Thr Pro Ile Gly
 1075 1080 1085

25 Leu Ile Ala Arg Asp Ser Pro His Val Leu Val Ala Leu Thr Arg His
 1090 1095 1100

Thr Lys Ala Met Val Tyr Tyr Thr Val Val Phe Asp Ala Val Thr Ser
 30 1105 1110 1115 1120

Ile Ile Ala Asp Val Glu Lys Val Asp Gln Ser Ile Leu Thr Met Phe
 1125 1130 1135

35 Ala Thr Thr Val Pro Thr Lys
 1140

40 <210> 19
 <211> 1503
 <212> DNA
 <213> Artificial Sequence

45 <220>
 <223> Description of Artificial Sequence: nucleotide
 sequence corresponding to 57 kD protein of CGMMV
 strain SH

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 gatttcgatg ccgtaaccat gcggttgagg gacaacgaat ttaacttaca acctgtagg 180
 ctaaccttga gtaatttaga tccggtaccc gctttgatta agaatgaagc gcagaatttt 240
 55 ctgatccccg ttttgcgtac ggctgtgaa aggcgcgcga ttccgggtct tcttgagaat 300
 ctttagtagta tgataaagag gaatatgaat actcctgatt tagctgggac cgtagatata 360
 actaacatgt cgatttctat agtagataac ttcttttctt cttttgttag ggacgaggtt 420
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 tcgtgtcaac caacctcagc ggttgccagc ttagctaatt tcaatttcat agatttgcct 540
 60 gcctttgata cttatatgca tatgattaag aggcaaccca agagtcggtt agatacttcg 600
 attcagtcctg aatatccggc cttgcaaact attgtttatc accctaaagt ggtaaatgca 660

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10 tacttctgcg ggaagtatat aattcaccat gccaacggct gtattgttta ccctgaccct 1260
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ctcgacgatg caatccaaga attatttccct aatgctgggg gttgcagttt tgtaattaat 1440
tgtttgtgta agtatattgag tgataagcgc cttttccgta gtctttacat agatgtctct 1500
15 aag 1503

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<210> 20

<211> 501

20 <212> PRT

<213> Artificial Sequence

<220>

25 <223> Description of Artificial Sequence: amino acid
sequence corresponding to 57 kD protein of CGMMV
strain SH

<400> 20

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30 Met Gln Asn Ser Leu Tyr Val His Arg Asn Ile Phe Leu Pro Val Ser
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Lys Thr Gly Phe Tyr Thr Asp Met Gln Glu Phe Tyr Asp Arg Cys Leu
    20          25          30

35 Pro Gly Asn Ser Phe Val Leu Asn Asp Phe Asp Ala Val Thr Met Arg
    35          40          45

Leu Arg Asp Asn Glu Phe Asn Leu Gln Pro Cys Arg Leu Thr Leu Ser
    50          55          60

40 Asn Leu Asp Pro Val Pro Ala Leu Ile Lys Asn Glu Ala Gln Asn Phe
    65          70          75          80

Leu Ile Pro Val Leu Arg Thr Ala Cys Glu Arg Pro Arg Ile Pro Gly
    85          90          95

Leu Leu Glu Asn Leu Val Ala Met Ile Lys Arg Asn Met Asn Thr Pro
    100         105         110

50 Asp Leu Ala Gly Thr Val Asp Ile Thr Asn Met Ser Ile Ser Ile Val
    115         120         125

Asp Asn Phe Phe Ser Ser Phe Val Arg Asp Glu Val Leu Leu Asp His
    130         135         140

55 Leu Asp Cys Val Arg Ala Ser Ser Ile Gln Ser Phe Ser Asp Trp Phe
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Ser Cys Gln Pro Thr Ser Ala Val Gly Gln Leu Ala Asn Phe Asn Phe
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```


500

5 <210> 21
 <211> 4932
 <212> DNA
 <213> Artificial Sequence

10 <220>
 <223> Description of Artificial Sequence: nucleotide
 sequence corresponding to 186 kD protein of CGMMV
 strain SH

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 catcaagaca gacgcccgaa aatgaatttt tctcgtgtgg tcagcacaga gcacaccagg 180
 cttgtaactg acgcgtatcc ggagttttcg attagcttta ccgccaccaa gaactctgta 240
 20 cactcccttg cgggtggtct gaggttctt gaattggaat atatgatgat gcaggtgccc 300
 tacggctcac cttgttatga catcggcggt aactatacgc agcacttggt caaaggtaga 360
 tcatatgtgc attgctgcaa tccgtgccta gatcttaagg atgttgcgag gaatgtgatg 420
 tacaacgata tgattacgca acatgtacag aggcacaagg gatcttgccg gtgcagacct 480
 cttccaactt tccagataga tgcattcagg aggtacgata gttctccctg tgcggtcacc 540
 25 tgttcagacg ttttccaaga gtgttcctat gattttggga gtggtaggga taatcatgca 600
 gtctcgttgc attcaatcta cgatatccct tattcttcga tcggacctgc tcttcatagg 660
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aagtatggtt acttctgcgg gaagtatata attcaccatg ccaacggctg tattgtttac 4680
cctgaccctt taaaattaat tagtaaatta ggtaataaga gtcttgtagg gtatgagcat 4740
gttgaggagt ttcttatatc tctcctcgac gttgctcata gtttgtttaa tgggtgcttat 4800
35 ttccatttac tcgacgatgc aatccacgaa ttatttccta atgctggggg ttgcagtttt 4860
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40 <210> 22
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 <212> PRT
 <213> Artificial Sequence

45 <220>
 <223> Description of Artificial Sequence: amino acid
 sequence corresponding to 186 kD protein of CGMMV
 strain SH

50 <400> 22
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55 Ser Gly Arg Asn Asn Leu Val Ser Gln Leu Ala Ser Lys Arg Val Tyr
 20 25 30

Asp Glu Ala Val Arg Ser Leu Asp His Gln Asp Arg Arg Pro Lys Met
 35 40 45

60 Asn Phe Ser Arg Val Val Ser Thr Glu His Thr Arg Leu Val Thr Asp
 50 55 60

	Ala	Tyr	Pro	Glu	Phe	Ser	Ile	Ser	Phe	Thr	Ala	Thr	Lys	Asn	Ser	Val	65	70	75	80
5	His	Ser	Leu	Ala	Gly	Gly	Leu	Arg	Leu	Leu	Glu	Leu	Glu	Tyr	Met	Met	85	90	95	
	Met	Gln	Val	Pro	Tyr	Gly	Ser	Pro	Cys	Tyr	Asp	Ile	Gly	Gly	Asn	Tyr	100	105	110	
10	Thr	Gln	His	Leu	Phe	Lys	Gly	Arg	Ser	Tyr	Val	His	Cys	Cys	Asn	Pro	115	120	125	
15	Cys	Leu	Asp	Leu	Lys	Asp	Val	Ala	Arg	Asn	Val	Met	Tyr	Asn	Asp	Met	130	135	140	
	Ile	Thr	Gln	His	Val	Gln	Arg	His	Lys	Gly	Ser	Cys	Gly	Cys	Arg	Pro	145	150	155	160
20	Leu	Pro	Thr	Phe	Gln	Ile	Asp	Ala	Phe	Arg	Arg	Tyr	Asp	Ser	Ser	Pro	165	170	175	
	Cys	Ala	Val	Thr	Cys	Ser	Asp	Val	Phe	Gln	Glu	Cys	Ser	Tyr	Asp	Phe	180	185	190	
25	Gly	Ser	Gly	Arg	Asp	Asn	His	Ala	Val	Ser	Leu	His	Ser	Ile	Tyr	Asp	195	200	205	
30	Ile	Pro	Tyr	Ser	Ser	Ile	Gly	Pro	Ala	Leu	His	Arg	Lys	Asn	Val	Arg	210	215	220	
	Val	Cys	Tyr	Ala	Ala	Phe	His	Phe	Ser	Glu	Ala	Leu	Leu	Leu	Gly	Ser	225	230	235	240
35	Pro	Val	Gly	Asn	Leu	Asn	Ser	Ile	Gly	Ala	Gln	Phe	Arg	Val	Asp	Gly	245	250	255	
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40	His	Ser	Leu	Glu	Asn	Ile	Lys	Leu	Ile	Val	Met	Arg	Thr	Tyr	Phe	Pro	275	280	285	
45	Ala	Asp	Asp	Arg	Phe	Val	Tyr	Ile	Lys	Glu	Phe	Met	Val	Lys	Arg	Val	290	295	300	
	Asp	Thr	Phe	Phe	Phe	Arg	Leu	Val	Arg	Ala	Asp	Thr	His	Met	Leu	His	305	310	315	320
50	Lys	Ser	Val	Gly	His	Tyr	Ser	Lys	Ser	Lys	Ser	Glu	Tyr	Phe	Ala	Leu	325	330	335	
	Asn	Thr	Pro	Pro	Ile	Phe	Gln	Asp	Lys	Ala	Thr	Phe	Ser	Val	Trp	Phe	340	345	350	
55	Pro	Glu	Ala	Lys	Lys	Val	Leu	Ile	Pro	Lys	Phe	Glu	Leu	Ser	Arg	Phe	355	360	365	
60	Leu	Ser	Gly	Asn	Val	Lys	Ile	Ser	Arg	Met	Leu	Val	Asp	Ala	Asp	Phe	370	375	380	

	Val	His	Thr	Ile	Ile	Asn	His	Ile	Ser	Thr	Tyr	Asp	Asn	Lys	Ala	Leu	
	385					390					395					400	
5	Val	Trp	Lys	Asn	Val	Gln	Ser	Phe	Val	Glu	Ser	Ile	Arg	Ser	Arg	Val	
				405						410					415		
	Ile	Val	Asn	Gly	Val	Ser	Val	Lys	Ser	Glu	Trp	Asn	Val	Pro	Val	Asp	
				420					425					430			
10	Gln	Leu	Thr	Asp	Ile	Ser	Phe	Ser	Ile	Phe	Pro	Leu	Val	Lys	Val	Arg	
			435					440						445			
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		450					455						460				
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				500					505					510			
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	Arg	Glu	Ser	Val	Ser	Glu	Leu	Leu	Ala	Ser	Gly	Asp	Asp	Leu	Phe	Lys	
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		690					695					700					
60	Arg	Leu	Ser	Asn	Met	Asn	Phe	Phe	Lys	Val	Ser	Asp	Leu	Lys	Leu	Lys	

	705				710					715					720	
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5	Met	Lys	Asn	Tyr	Ile	Asp	Tyr	Leu	Ser	Ala	Ser	Leu	Gly	Ser	Thr	Leu
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		770					775					780				
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	785					790					795					800
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25	Thr	Lys	Val	Tyr	Ser	Val	Ile	Arg	Ser	Leu	Glu	Val	Leu	Asn	Lys	Glu
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	Asp	Leu	Val	Leu	Thr	Pro	Gly	Arg	Glu	Ala	Ala	Ala	Met	Ile	Arg	Arg
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40	Arg	Thr	Phe	Asp	Ser	Phe	Val	Met	Asn	Arg	Lys	Ile	Phe	Lys	Phe	Asp
			915					920					925			
	Ala	Val	Tyr	Val	Asp	Glu	Gly	Leu	Met	Val	His	Thr	Gly	Leu	Leu	Asn
		930					935					940				
45	Phe	Ala	Leu	Lys	Ile	Ser	Gly	Cys	Lys	Lys	Ala	Phe	Val	Phe	Gly	Asp
	945					950					955					960
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			995					1000					1005			
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 5 Lys Ser Gly Tyr Asn Asp Val Asn Thr Val His Glu Ile Gln Gly Glu
 1060 1065 1070
 Thr Phe Glu Glu Thr Ala Val Val Arg Ala Thr Pro Thr Pro Ile Gly
 1075 1080 1085
 10 Leu Ile Ala Arg Asp Ser Pro His Val Leu Val Ala Leu Thr Arg His
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 1105 1110 1115 1120
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 1395 1400 1405
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 1410 1415 1420
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 1425 1430 1435 1440
 15 Asp Asp Ile Leu Ala Trp Met Trp Ser Met Gly His Lys Arg Thr Ile
 1445 1450 1455
 20 Leu Gln Asp Phe Gln Ala Gly Ile Lys Thr Leu Ile Tyr Tyr Gln Arg
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 1475 1480 1485
 25 Ala Cys Val Ala Ser Met Leu Pro Leu Asp Lys Cys Phe Lys Ala Ser
 1490 1495 1500
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 1505 1510 1515 1520
 30 Pro Asp Ile Gln Ala Thr Ala Asn Leu Val Trp Asn Phe Glu Ala Lys
 1525 1530 1535
 35 Leu Phe Arg Lys Lys Tyr Gly Tyr Phe Cys Gly Lys Tyr Ile Ile His
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 His Ala Asn Gly Cys Ile Val Tyr Pro Asp Pro Leu Lys Leu Ile Ser
 1555 1560 1565
 40 Lys Leu Gly Asn Lys Ser Leu Val Gly Tyr Glu His Val Glu Glu Phe
 1570 1575 1580
 Arg Ile Ser Leu Leu Asp Val Ala His Ser Leu Phe Asn Gly Ala Tyr
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 45 Phe His Leu Leu Asp Asp Ala Ile His Glu Leu Phe Pro Asn Ala Gly
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 <211> 1139
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<220>

<223> Description of Artificial Sequence: nucleotide
sequence corresponding to sequence encoding coat
protein of CGMMV strain SH

5

<400> 23

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30

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<223> Description of Artificial Sequence: primer 97G01

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24

40

<210> 25

<211> 17

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<213> Artificial Sequence

45

<220>

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<400> 25

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17

50

<210> 26

<211> 21

<212> DNA

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<213> Artificial Sequence

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<223> Description of Artificial Sequence: primer 97G03

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<210> 35
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50 <210> 36
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<210> 37
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<223> Description of Artificial Sequence: primer 98L103

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<210> 38
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<210> 39
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<210> 41
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sequence 97G41

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<212> DNA
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20 <223> Description of Artificial Sequence: synthetic
sequence 97G42

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30 <210> 44
<211> 30
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic
sequence 97G43
35

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